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WORLD INTELLECTUAL PROPERTY ORGANIZATION

INTERNATIONAL APPLICATION PUBLISHED



WO 9604391A1

(51) International Patent Classification 6 :

C12N 15/64, 15/67, 15/85, 9/72, 5/10

A1

(43) International Publication Date: 15 February 1996 (15.02.96)

(21) International Application Number: PCT/US95/09576

(22) International Filing Date: 28 July 1995 (28.07.95)

(30) Priority Data:
08/286,740 5 August 1994 (05.08.94) US

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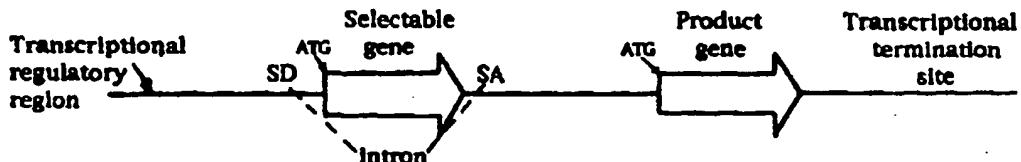
(81) Designated States: AU, CA, JP, MX, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).

Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS



(57) Abstract

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing host cells, a method of producing a protein of interest in high yields and a method of producing eukaryotic cells having multiple copies of a sequence encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells in a functional form has provided the key to understanding many fundamental biological processes, and has made possible the production of important proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several common problems exist that may limit the efficiency with which a gene encoding a desired protein can be introduced into and expressed in a host cell. One problem is knowing when the gene has been successfully transferred into recipient cells. A second problem is distinguishing between those cells that contain the gene and those that have survived the transfer procedures but do not contain the gene. A third problem is identifying and isolating those cells that contain the gene and that are expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic cells tend to be highly inefficient. Of the cells in a given culture, only a small proportion take up and express exogenously added DNA, and an even smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene encoding a desired protein typically is achieved by introducing into the same cells another gene, commonly referred to as a selectable gene, that encodes a selectable marker. A selectable marker is a protein that is necessary for the growth or survival of a host cell under the particular culture conditions chosen, such as an enzyme that confers resistance to an antibiotic or other drug, or an enzyme that compensates for a metabolic or catabolic defect in the host cell. For example, selectable genes commonly used with eukaryotic cells include the genes for aminoglycoside phosphotransferase (APH), hygromycin phosphotransferase (hyg), dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin, glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene on the basis of expression by the host cell of a second incorporated gene encoding a selectable marker is referred to as cotransfection (or cotransfected). In that method, a gene encoding a desired polypeptide and a selection gene typically are introduced into the host cell simultaneously, although they may be introduced sequentially. In the case of simultaneous cotransfection, the gene encoding the desired polypeptide

and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by 5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency 10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter 15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and 20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

25 Enhancers stimulate transcription from a linked promoter. Unlike promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters listed above also contain strong enhancers. Bendig, Genetic Engineering, 30 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing 35 the gene copy number in the host cell. One method for obtaining high gene copy number is to directly introduce into the host cell multiple copies of the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransflection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high 40 copy number. Furthermore, because no generally applicable, convenient method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning 45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors

derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective 10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under 15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell 20 is cotransfected with a product gene encoding a desired protein and a DHFR gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are 25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic 30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant 40 DHFR gene that encodes a protein with reduced sensitivity to methotrexate may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a neor gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the 5 product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the 10 selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of 15 transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 20 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such 25 recombinant host cells in a single step. One strategy involves co-transfected host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. 30 Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production 35 processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. 40 Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors 45 exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320 [1988]; Jang et al., J. Virol., 63:1651 [1989]).

A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate 5 transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10

SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the 15 selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates 20 expression of the product gene using the transcriptional regulatory region.

In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

25

In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time 30 for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

35

After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high 40 levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines 5 shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various 10 splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence (Δ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 15 3 used for expression of anti-IgE V_H.

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron-(WT ras SD)-tPA expression vector of Example 1.

20 Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [Δ GT]) and 25 from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 30 Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the Δ GT pool) were exposed to 200nM Mtx to select for 35 DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron-(WT ras SD)-TNFr-IgG expression vector of Example 2.

40 Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12. CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

45 Figure 8 depicts schematically the DNA construct used for expression of the V_L of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V_H expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V_L expression vector of Example 3.

5 Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V_H) and light (V_L) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

10 DESCRIPTION OF THE PREFERRED EMBODIMENTS

Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the 15 chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of 20 growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in Enzymology, 185: 537-566 (1990), for a review of these.

TABLE 1
Selectable Genes and their Selection Agents

	Selection Agent	Selectable Gene
	Methotrexate	Dihydrofolate reductase
30	Cadmium	Metallothionein
	PALA	CAD
	Xyl-A-or adenosine and 2'-deoxycoformycin	Adenosine deaminase
35	Adenine, azaserine, and coformycin	Adenylylate deaminase
	6-Azauridine, pyrazofuran	UMP Synthetase
	Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs e.g. adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	β -Aspartyl hydroxamate or Albizzini	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	α -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na'K'-ATPase

- The preferred selectable gene is an amplifiable gene. As used herein, 20 the term "amplifiable gene" refers to a gene which is amplified (i.e. additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (i.e. an amplifiable marker) which is required for growth of eukaryotic cells under those conditions.
- 25 For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mt_x. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko et al., *supra*).
- 30 As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary 35 nutrient solutions. In addition, any of the media described in Ham and Wallace, *Meth. Enz.*, 58:44 (1979), Barnes and Sato, *Anal. Biochem.*, 102:255

(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be 5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as GentamycinTM drug), trace elements (defined as inorganic compounds usually 10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with 15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table 20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA 25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a 30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a cis-acting DNA element, 35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the 40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product 45 genes suitably encode a peptide, or may encode a polypeptide sequence of

amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and β -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- β ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- β 1, TGF- β 2, TGF- β 3, TGF- β 4, or TGF- β 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- β , TGF- α , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- γ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to 5 translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or de novo synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gatermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, 15 or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 20 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 25 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor 30 sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer 35 to the *in vitro* amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone 40 specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA 45 samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced
5 transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the
10 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett,
et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence
15 immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a
20 nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice
25 acceptor site occurs within the consensus sequence (U/C)₁₁NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host
30 cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

35 The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by
40 northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are
45 independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability
5 and/or copy number of a transcribed sequence in order to allow for elevated
levels of a RNA sequence of interest. In general, the methods of the
present invention involve transfected a eukaryotic host cell with an
expression vector comprising both a product gene encoding a desired
polypeptide and a selectable gene (preferably an amplifiable gene).

10 Selectable genes and product genes may be obtained from genomic DNA,
cDNA transcribed from cellular RNA, or by in vitro synthesis. For example,
libraries are screened with probes (such as antibodies or oligonucleotides
of about 20-80 bases) designed to identify the selectable gene or the
product gene (or the protein(s) encoded thereby). Screening the cDNA or
15 genomic library with the selected probe may be conducted using standard
procedures as described in chapters 10-12 of Sambrook et al., Molecular
Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory
Press, 1989). An alternative means to isolate the selectable gene or
product gene is to use PCR methodology as described in section 14 of
20 Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully
selected oligonucleotide sequences to screen cDNA libraries from various
tissues known to contain the selectable gene or product gene. The
oligonucleotide sequences selected as probes should be of sufficient length
25 and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected
upon hybridization to DNA in the library being screened. The preferred
method of labeling is to use ³²P-labeled ATP with polynucleotide kinase,
as is well known in the art, to radiolabel the oligonucleotide. However,
30 other methods may be used to label the oligonucleotide, including, but not
limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is
preceded by DNA encoding a signal sequence having a specific cleavage site
at the N-terminus of the mature protein or polypeptide. In general, the
35 signal sequence may be a component of the expression vector, or it may be
a part of the selectable gene or product gene that is inserted into the
expression vector. If a heterologous signal sequence is used, it
preferably is one that is recognized and processed (i.e., cleaved by a
signal peptidase) by the host cell. For yeast secretion the native signal
40 sequence may be substituted by, e.g., the yeast invertase leader, alpha
factor leader (including *Saccharomyces* and *Kluyveromyces* α-factor leaders,
the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or
acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179
published 4 April 1990), or the signal described in WO 90/13646 published
45 15 November 1990. In mammalian cell expression the native signal sequence

of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spiced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfecants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as *de novo* synthesis or *in vitro* deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general 5 proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction 10 endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single 15 transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., *J. Biol. Chem.*, 255:2073 [1980]) or other glycolytic enzymes (Hess et al., *J. Adv. Enzyme Reg.*, 7:149 [1968]; and Holland, 20 *Biochemistry*, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the 25 additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytchrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and 30 promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all 35 eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as 40 polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters, 45 and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a HindIII E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; Reyes et al., Nature, 297:598-601 (1982) on expression of human β -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon β 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA

fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 μ plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in 15 *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, 20 and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for 25 cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic 30 host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvcourt et al., J. Bacteriol., 737 (1983)], *yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., 35 Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

40 Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and 45 insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*

(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda* cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen., 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dpi2. CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly ³²P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescens, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay 5 of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels, 10 fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is 15 measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human 20 origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged 25 to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate 30 precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and 35 literature references cited herein are expressly incorporated by reference.

EXAMPLE 1

tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such 40 as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the 45 cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V_h) gene (Eaton et al., Biochem., 25:8343 [1986]).

5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that comprises the splice donor site. Those sequences were altered by 10 overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control (Δ GT). A polylinker was 25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site (Δ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease 35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 μ l 1/10 Tris EDTA. Then, 10 μ g of DNA was incubated with 10 7 CHO.dp12 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before 40 electroporation at 400 volts and 330 μ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

45 To prepare the mRNA, RNA was extracted from 5 x 10 7 cells which were grown from pools of more than 200 clones derived from the stable

transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10 μ g of mRNA was then subjected to Northern blotting which involved running the mRNA on 5 a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and 10 spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only 15 full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after 20 duplicate electroporations with the three intron vectors described above and from a conventional vector that has a CMV promoter driving tPA and a SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector 25 efficiently generates colonies and does not vary significantly from the Δ GT construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluence. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was 30 coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling 35 the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open 40 reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors 45 (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA. SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing (Δ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2 μ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR^r cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

EXAMPLE 2

TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., *Proc. Natl. Acad. Sci. USA*, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site (Δ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 $\mu\text{g}/\text{ml}$ when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, 5 contrary to expectations, it was discovered that isolation of high product expressing host DHFR^r cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

EXAMPLE 3

Antibody production using a dicistronic expression vector

10 The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the 15 promoter/enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNFr-IgG constructs. 20 Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the 25 vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream 30 of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression. 35 All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was 40 sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1 μM and achieved a greater than 2 fold increase in antibody titer. The 1 μM Mtx 45 resistant pool achieved a titer of 41 mg/L when grown under optimal conditions in suspension culture.

The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S³⁵ cysteine and 5 methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50 μ Ci each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run 10 on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction 15 the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain 20 expressed by the pool.

CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and 25 a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras 30 gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number 35 is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNFr immunoadhesin or anti-IgE V_H.

The homogeneity of expression of clones generated by the ras splice 40 donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived 45 from the conventional vector may be the result of breaks in the tPA or

TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or 5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased 10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely 15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain 25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain 30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 μ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly 45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- 10 (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 15 (iii) NUMBER OF SEQUENCES: 4
- 20 (iv) CORRESPONDENCE ADDRESS:
 (A) ADDRESSEE: Genentech, Inc.
 (B) STREET: 460 Point San Bruno Blvd
 (C) CITY: South San Francisco
 (D) STATE: California
 (E) COUNTRY: USA
 (F) ZIP: 94080
- 25 (v) COMPUTER READABLE FORM:
 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
 (B) COMPUTER: IBM PC compatible
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 (D) SOFTWARE: patin (Genentech)
- 30 (vi) CURRENT APPLICATION DATA:
 (A) APPLICATION NUMBER:
 (B) FILING DATE:
 (C) CLASSIFICATION:
- 35 (vii) PRIOR APPLICATION DATA:
 (A) APPLICATION NUMBER: 08/286740
 (B) FILING DATE: 05-AUG-1994
- 40 (viii) ATTORNEY/AGENT INFORMATION:
 (A) NAME: Lee, Wendy M.
 (B) REGISTRATION NUMBER: 00,000
 (C) REFERENCE/DOCKET NUMBER: 798PCT
- 45 (ix) TELECOMMUNICATION INFORMATION:
 (A) TELEPHONE: 415/225-1994
 (B) TELEFAX: 415/952-9881
 (C) TELEX: 910/371-7168
- 50 (2) INFORMATION FOR SEQ ID NO:1:
- 55 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 7360 bases
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- 60 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50

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70 TTACGGTAAA TGCCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150

75 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200

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 55 GAAGGAGCTA ACCGCTTTTG TGCAACACAT GGGGGATCAT GTAACTCGCC 5800
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 60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900
 TGGCGAACTA CTTACTCTAG CTTCCGGCA ACAATTAATA GACTGGATGG 5950
 65 AGGCGGATAA AGTTGCAGGA CCACCTCTGC GCTCGGCCCT TCCGGCTGGC 6000

TGGTTTATTG CTGATAAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050
5 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100
ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150
10 GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTA 6200
CTCATATATA CTTTAGATTG ATTTAAAATC TCATTTTAA TTTAAAAGGA 6250
15 TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAAT CCCTTAACGT 6300
GAGTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350
20 TTCTTGAGAT CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA 6400
AACACCAGCT ACCAGCGGTG GTTGTGTTGC CGGATCAAGA GCTACCAACT 6450
CTTTTCCGA AGGTAACTGG CTTCAGCAGA GCGCAGATAAC CAAATACTGT 6500
30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550
CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCACTGGC TGCTGCCAGT 6600
35 GGCGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650
TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA CAGCCCAGCT 6700
40 TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750
GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800
50 CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGGAAACG 6850
CCTGGTATCT TTATAGTCCT GTGGGTTTC GCCACCTCTG ACTTGAGCGT 6900
55 CGATTTTGT GATGCTCGTC AGGGGGCGG AGCCTATGGA AAAACGCCAG 6950
CAACCGGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA 7000
60 TGTTCTTCC TGCCTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC 7050
TTTGAGTGAG CTGATAACCGC TCGCCGGAGC CGAACGACCG AGCGCAGCGA 7100
65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCA TAATCCAGCT GGCACGACAG GTTCCCCAC 7200

5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250

TTAGGCACCC CAGGCTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300

10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350

TACGAATTAA 7360

15

(2) INFORMATION FOR SEQ ID NO:2:

- 20 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 6889 bases
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50

30 TACGGGGTCA TTAGTTCATATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100

35 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150

ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200

40 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCAC TTGGCAGTAC 250

ATCAAGTGTAA TCATATGCCA AGTACGCCCT CTATTGACGT CAATGACGGT 300

45 AAATGGCCCG CCTGGCATTAA TGCCCAAGTAC ATGACCTTAT GGGACTTTCC 350

50 TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400

GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450

55 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500

AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550

60 AAATGGGCGG TAGGCGTGTAA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

65 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750
5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTCGA CCATTGAACT 800
GCATCGTCGC CGTGTCCAA AATATGGGA TTGGCAAGAA CGGAGACCTA 850
10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900
AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAA 950
15 CCTGGTTCTC CATTCTGAG AAGAATCGAC CTTAAAGGA CAGAATTAAT 1000
ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTCT 1050
20 TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100
25 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAAG 1150
GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTGTGA CAAGGATCAT 1200
30 GCAGGAATTG GAAAGTGACA CGTTTTCCC AGAAATTGAT TTGGGGAAAT 1250
ATAAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300
35 AAAGGCATCA AGTATAAGTT TGAAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350
40 AGATGCTTTC AAGTTCTCTG CTCCCCCTCCT AAAGCTATGC ATTTTTATAA 1400
GACCATGGGA CTTTGCTGG CTTTAGACCC CCTTGGCTTC GTTAGAACGC 1450
45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500
50 CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACTCCA 1550
GGTCAACTGC ACCTCGGTTC TATCGATTGA ATCCCCGGC CATAGCTGTC 1600
55 TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650
TGGAGCTGTT GGTGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT 1700
60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750
TATCCACCCCT CAAAATAATT CGATTTGCTG TACCAAGTGC CACAAAGGAA 1800
65 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGACTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTC AAAAAACCAC TCAGACACTG 1900
5 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950
CTTGCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000
10 CGGCATTATT GGAGTGAAA CCTTTTCCAG TGCTTCATT GCAGCCTCTG 2050
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAACAG AACACCGTGT 2100
15 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150
AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCCAGAT 2200
20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250
25 AGCTCAAAC CCCACTTGGT GACACAACTC ACACATGCC ACGGTGCCCCA 2300
GAGCCAAAT CTTGTGACAC ACCTCCCCCG TGCCCACGGT GCCCAGAGCC 2350
30 CAAATCTTGT GACACACCTC CCCCATGCC ACGGTGCCCCA GAGCCAAAT 2400
CTTGTGACAC ACCTCCCCCA TGCCCACGGT GCCCAGCACC TGAACCTCTG 2450
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCA AAACCCAAGG ATACCCATTAT 2500
40 GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550
AAGACCCCGA GGTCCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT 2600
45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650
GGTCAGCGTC CTCACCGTCC TGCAACCAGGA CTGGCTGAAC GGCAAGGAGT 2700
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCAT CGAGAAAACC 2750
55 ATCTCCAAA CCAAAGGACA GCCCGAGAA CCACAGGTGT ACACCCCTGCC 2800
CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTCAGCCTG ACCTGCCTGG 2850
60 TCAAAGGCTT CTACCCCAGC GACATGCCG TGGAGTGGGA GAGCAGCGGG 2900
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950
65 CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC 3000

AGGGGAACAT CTTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC 3050
5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAAATGAG TGCGACGGCC 3100
GGGGATCCTC TAGAGTCGAC CTGCAGAACG TTGGCCGCCA TGGCCCAACT 3150
10 TGTTTATTGC AGCTTATAAT GGTTACAAAT AAAGCAATAG CATCACAAAT 3200
TTCACAAATA AAGCATTTC TTCACTGCAT TCTAGTTGTG GTTGTCCAA 3250
15 ACTCATCAAT GTATCCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300
GCAGCACCAT GCCCTGAAAT AACCTCTGAA AGAGGAACCTT GGTTAGGTAC 3350
20 CTTCTGAGGC GGAAAGAACCG AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400
25 GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450
AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500
30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550
TAACTCCGCC CATCCC GCCCT CTAACTCCGC CCAGTTCCGC CCATTCTCCG 3600
35 CCCCATGGCT GACTAATTTT TTTTATTAT GCAGAGGCCG AGGCCGCCCTC 3650
40 GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTT GGAGGCCTAG 3700
GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTACAAC 3750
45 GTCGTGACTG GGAAAACCCCT GGCGTTACCC AACTTAATCG CCTTGCAGCA 3800
CATCCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCC GCACCGATCG 3850
50 CCCCTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900
55 ATTTTCTCCT TACGCATCTG TGCGGTATTT CACACCGCAT ACGTCAAAGC 3950
AACCATAGTA CGCGCCCTGT AGCGGCCAT TAAGCGCGGC GGGTGTGGTG 4000
60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050
TTTCGCTTTC TTCCCTTCCT TTCTCGCCAC GTTCGCCGGC TTTCCCCGTC 4100
65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAAACT TGATTTGGGT GATGGTCAC GTAGTGGCC 4200
5 ATCGCCCTGA TAGACGGTTT TTGCCCCTTT GACGTTGGAG TCCACGTTCT 4250
TTAATAGTGG ACTCTTGTTTC CAAACTGGAA CAACACTCAA CCCTATCTG 4300
10 GGCTATTCTT TTGATTTATA AGGGATTTG CCGATTTCGG CCTATTGGTT 4350
AAAAAATGAG CTGATTTAAC AAAAATTAA CGCGAATTAA AACAAAATAT 4400
15 TAACGTTTAC AATTTTATGG TGCACCTCTCA GTACAATCTG CTCTGATGCC 4450
GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500
20 CCCCGACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550
25 CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600
GTCAGAGGTT TTCACCGTCA TCACCGAAAC GCGCGAGGCA GTATTCTTGA 4650
30 AGACGAAAGG GCCTCGTGAT ACGCCTATT TTATAGGTTA ATGTCATGAT 4700
ATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGA AATGTGCGCG 4750
35 GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC 4800
40 ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG 4850
TATGAGTATT CAACATTTCC GTGTCGCCCT TATTCCCTTT TTTGCGGCAT 4900
45 TTTGCCTTCC TGTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT 4950
GCTGAAGATC AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA 5000
50 CAGCGGTAAG ATCCTTGAGA GTTTTCGCC CGAAGAACGT TTTCAATGA 5050
55 TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100
GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT 5150
60 GGTTGAGTAC TCACCAAGTCA CAGAAAAGCA TCTTACGGAT GGCGATGACAG 5200
TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC 5250
65 AACTTACTTC TGACAAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTT 5300

GCACAAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350
5 TGAATGAAGC CATAACAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400
ATGGCAACAA CGTTGCGCAA ACTATTAACG GGCGAACTAC TTACTCTAGC 5450
10 TTCCCCGCAA CAATTAATAG ACTGGATGGA GGCGGATAAA GTTGCAGGAC 5500
CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGTTTATTGC TGATAAAATCT 5550
15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600
TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA 5650
20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700
25 AAGCATTGGT AACTGTCAGA CCAAGTTAAC TCATATATAC TTTAGATTGA 5750
TTTAAAACCTT CATTTTTAAT TTAAAAGGAT CTAGGTGAAG ATCCTTTTG 5800
30 ATAATCTCAT GACCAAAATC CCTTAACGTG AGTTTCGTT CCACTGAGCG 5850
TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTCT 5900
35 GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG 5950
40 TTTGTTGCC GGATCAAGAG CTACCAACTC TTTTTCCGAA GGTAACTGGC 6000
TTCAGCAGAG CGCAGATAACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT 6050
45 AGGCCACAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100
TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150
50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTCGGGCTG 6200
55 AACGGGGGGT TCGTGCACAC AGCCCAGCTT GGAGCGAACG ACCTACACCG 6250
AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA 6300
60 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAAGGTG GAACAGGAGA 6350
GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400
65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTTGTG ATGCTCGTCA 6450

GGGGGCGGGA GCCTATGGAA AAACGCCAGC AACGCCCTT TTTACGGTT 6500
 5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTCCT GCGTTATCCC 6550
 CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT 6600
 10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650
 AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTCAATT 6700
 15 AATCCAGCTG GCACGACAGG TTTCGGACT GGAAAGCGGG CAGTGAGCGC 6750
 AACGCAATTAA ATGTGAGTTA CCTCACTCAT TAGGCACCCCC AGGCTTTACA 6800
 20 CTTTATGCTT CCGGCTCGTA TGGTGTGTGG AATTGTGAGC GGATAACAAT 6850
 25 TTCACACAGG AAACAGCTAT GACCATGATT ACGAATTAA 6889

(2) INFORMATION FOR SEQ ID NO:3:

30 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6557 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50
 GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100
 45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150
 TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTAA 200
 50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250
 55 CGCCCAGTTC CGCCCATTCT CCGCCCCATG GCTGACTAAT TTTTTTTATT 300
 TATGCAGAGG CCGAGGCCGC CTCGGCTCT GAGCTATTCC AGAAGTAGTG 350
 60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAAGCTA GCTTATCCGG 400
 65 CCGGGAACGG TGCATTGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450
 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500

TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA 550
5 AGAACGGAGA CCTACCCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600
CAAAGAACATGA CCACAACCTC TTCAGTGAA GGTAACAGA ATCTGGTGT 650
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750
15 GGAGCTCATT TTCTTGCCAA AAGTTGGAT GATGCCTTAA GACTTATTGA 800
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTGGATA GTCGGAGGCA 850
20 GTTCTGTTA CCAGGAAGCC ATGAATCAC CAGGCCACCT TAGACTCTTT 900
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950
TGATTGGGG AAATATAAAC CTCTCCAGA ATACCCAGGC GTCCTCTCTG 1000
30 AGGTCCAGGA GGAAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100
35 ATGCATTTT ATAAGACCAT GGGACTTTG CTGGCTTTAG ATCCCCTTGG 1150
40 CTTCGTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATAACAC 1200
ATACGATTAA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGGTTCTAT CGATTGAATT 1300
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350
50 GGAGTACATT CAGAAGTTCA GCTGGGGAG TCTGGCGGTG GCCTGGTGCA 1400
55 GCCAGGGGGC TCACTCCGTT TGTCCGTGTC AGTTTCTGGC TACTCCATCA 1450
CCTCCGGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500
60 GAATGGGTTG CATCGATTAC GTATGCCGGA TCGACTAACT ATAACCCTAG 1550
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCGT CTATTATTGT 1650

GCTCGAGGCA GCCACTATTT CGGCGCCTGG CACTTCGCCG TGTGGGTCA 1700
5 AGGAACCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750
10 TCCCCCTGGC ACCCTCCCTCC AAGAGCACCT CTGGGGCAC AGCGGCCCTG 1800
15 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCGTGGAA 1850
20 CTCAGGCGCC CTGACCAGCG GCGTGCACAC CTTCCCGCT GTCCTACAGT 1900
25 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGACTGTGCC CTCTAGCAGC 1950
30 TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGAACAC 2000
35 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050
40 GCCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCCTC 2100
45 TTCCCCCCAA AACCCAAGGA CACCCTCATG ATCTCCCGA CCCCTGAGGT 2150
50 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCGTAG GTCAAGTTCA 2200
55 ACTGGTACGT GGACGGCGTG GAGGTGCATA ATGCCAAGAC AAAGCCGCGG 2250
60 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300
65 GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350
70 AAGCCCTCCC AGCCCCCATC GAGAAAACCA TCTCCAAAGC CAAAGGGCAG 2400
75 CCCCAGAAC CACAGGTGTA CACCCCTGCC CCATCCCGGG AAGAGATGAC 2450
80 CAAGAACCAAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCCAGCG 2500
85 ACATGCCGT GGAGTGGGAG AGCAATGGGC AGCCGGAGAA CAACTACAAG 2550
90 ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600
95 GCTCACCGTG GACAAGAGCA GGTGGCAGCA GGGGAACGTC TTCTCATGCT 2650
100 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700
105 CTGTCTCCGG GTAAATGAGT GCGACGGCCC TAGAGTCGAC CTGCAGAAGC 2750
110 TTGGCCGCCA TGGCCCAACT TGTTATTGC AGCTTATAAT GGTTACAAAT 2800

AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTT TTCACTGCAT 2850
5 TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGGAT 2900
CGATCGGGAA TTAATTGGC GCAGCACCAT GGCTGAAAT AACCTCTGAA 2950
10 AGAGGAACCTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000
TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA 3050
15 GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC 3100
CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC 3150
20 AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAACTCCGC 3200
25 CCAGTTCCGC CCATTCTCCG CCCCCATGGCT GACTAATTTT TTTTATTAT 3250
GCAGAGGCCG AGGCCGCCTC GGCTCTGAG CTATTCCAGA AGTAGTGAGG 3300
30 AGGCTTTTTT GGAGGCCTAG GCTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350
CCGCTTAATT AAGGCGCGCC ATTTAAATCC TGCAAGTAAC AGCTTGGCAC 3400
35 TGGCCGTCGT TTTACAACGT CGTGACTGGG AAAACCTGG CGTTACCCAA 3450
40 CTTAATCGCC TTGCAGCACA TCCCCCTTC GCCAGCTGGC GTAATAGCGA 3500
AGAGGCCCGC ACCGATCGCC CTTCCAACA GTTGCCTAGC CTGAATGGCG 3550
45 AATGGCGCCT GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTTCA 3600
CACCGCATAAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT 3650
50 AGCGCGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG 3700
55 CGCCCTAGCG CCCGCTCCTT TCGCTTCTT CCCTTCCTT CTCGCCACGT 3750
TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC 3800
60 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAAACTTG ATTTGGGTGA 3850
TGGTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTGA 3900
65 CGTTGGAGTC CACGTTCTT AATAGTGGAC TCTTGTCCA AACTGGAACA 3950

ACACTCAACC CTATCTGGG CTATTCTTT GATTTATAAG GGATTTGCC 4000
5 GATTTCGGCC TATTGGTAA AAAATGAGCT GATTTAACAA AAATTTAACG 4050
CGAATTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100
10 ACAATCTGCT CTGATGCCGC ATAGTTAACG CAACTCCGCT ATCGCTACGT 4150
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250
TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC 4300
20 20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTT 4350
25 ATAGGTTAAT GTCAATGATAA TAATGGTTTC TTAGACGTCA GGTGGCACTT 4400
TTCGGGGAAA TGTGGCGGA ACCCCTATTG GTTTATTTT CCAAATACAT 4450
30 TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCATAA 4500
ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTTCCGT GTGCCCTTA 4550
35 TTCCCTTTTG TCGGCATTT TGCCTTCCTG TTTTGCTCA CCCAGAAACG 4600
40 CTGGTGAAAG TAAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA 4650
CATCGAACTG GATCTCAACA GCGGTAAGAT CCTTGAGAGT TTTCGCCCCG 4700
45 AAGAACGTTT TCCAATGATG AGCACTTTA AAGTTCTGCT ATGTGGCGCG 4750
GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA 4800
50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATE 4850
TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG 4900
55 AGTGATAACA CTGCGGCCAA CTTACTTCTG ACAACGATCG GAGGACCGAA 4950
60 GGAGCTAACG GCTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050
65 ACCACGATGC CAGCAGCAAT GGCAACAAACG TTGCGCAAAC TATTAACCTGG 5100

CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150
5 CGGATAAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200
TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT 5250
10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300
CGACGGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA GATCGCTGAG 5350
15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400
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20 AGGTGAAGAT CCTTTTGAT AATCTCATGA CCAAAATCCC TTAACGTGAG 5500
25 TTTTCGTTCC ACTGAGCGTC AGACCCGTA GAAAAGATCA AAGGATCTTC 5550
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30 CACCGCTACC AGCGGTGGTT TGTTGCCGG ATCAAGAGCT ACCAACTCTT 5650
TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCTT 5700
35 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5750
40 CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800
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65 TCTTTCTGC GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CGCGAGCCGA ACGACCGAGC GCAGCGAGTC 6300

5 AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350

CGCGTTGGCC GATTCAATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400

10 AAAGCGGGCA GTGAGCGCAA CGCAATTAAAT GTGAGTTACC TCACTCATTAA 6450

GGCACCCAG GCTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA 6500

15 TTGTGAGCGG ATAACAATTACACACAGGAA ACAGCTATGA CCATGATTAC 6550

20 GAATTAA 6557

(2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7305 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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TACGGGGTCA TTAGTTCATATA CCCATATAT GGAGTTCCGC GTTACATAAC 100

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ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200

45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCAC TTGGCAGTAC 250

50 ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300

AAATGGCCCG CCTGGCATTAA TGCCCAAGTAC ATGACCTTAT GGGACTTTCC 350

55 TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400

GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTGA CTCACGGGA 450

60 TTTCCAAGTC TCCACCCCAT TGACGTCAAAT GGGAGTTGT TTTGGCACCA 500

AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550

65 AAATGGGCGG TAGGCGTGTAA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

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5 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG AACGGTGCA 700
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10 GTCTATAGGC CCACCCCCCTT GGCTTCGTTA GAACGCGGCT ACAATTAATA 800
CATAACCTTA TGTATCATAc ACATACGATT TAGGTGACAC TATAGAATAA 850
15 CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACCTCCAG GTCCAAC TGC 900
ACCTCGGTTA TAAGCTTATC GATATGAAAA AGCCTGAACT CACCGCGACG 950
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25 GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050
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30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150
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35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCC 1250
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50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGACTGCCCGA AAAGTCCGGC 1500
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5 ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATGCC 1850
CGCAGAACGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900
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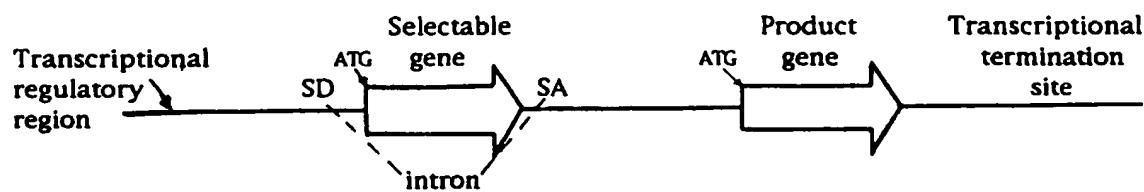
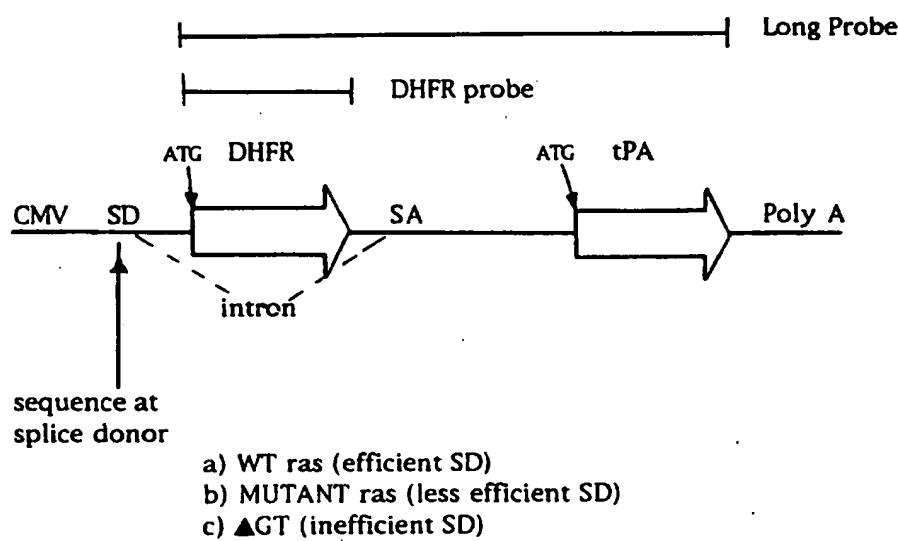
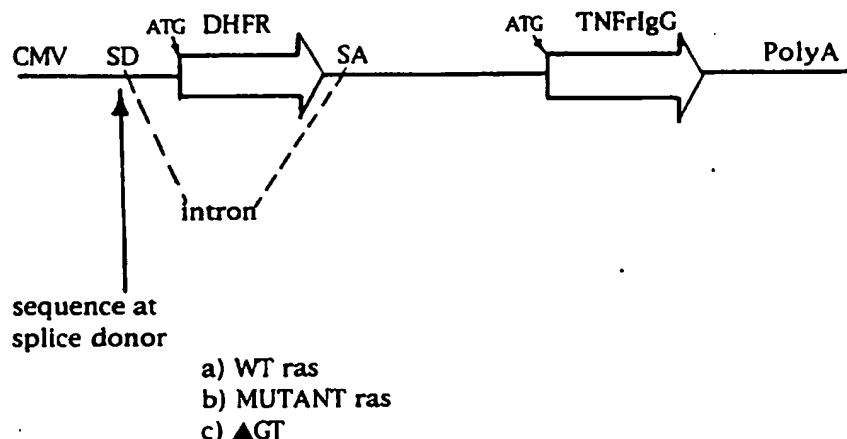
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40 GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050
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55 GTGAGCGGAT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300
ATTAA 7305
60

CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional regulatory region.
5
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.
20
5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.
25
7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.
30
9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic host.
35
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.
40
13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1 integrated into a chromosome of the host cell.
45

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the host cell of claim 11 so as to express the product gene and recovering the product from the host cell culture.
5
17. The method of claim 16 further comprising recovering the product from the culture medium.
- 10 18. The method of claim 16 wherein the selectable gene is an amplifiable gene and the splice donor site comprises an efficient splice donor sequence.
- 15 19. A method for producing a product of interest comprising culturing the host cell of claim 12 so as to express the product gene in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, and recovering the product.
- 20 20. A method for producing eukaryotic cells having multiple copies of a product gene comprising transforming eukaryotic cells with the DNA construct of claim 5, growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene.
25
21. The method of claim 20 further comprising recovering from the selected cells the product of interest.

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FIG. 1A**FIG. 1B****FIG. 1C**

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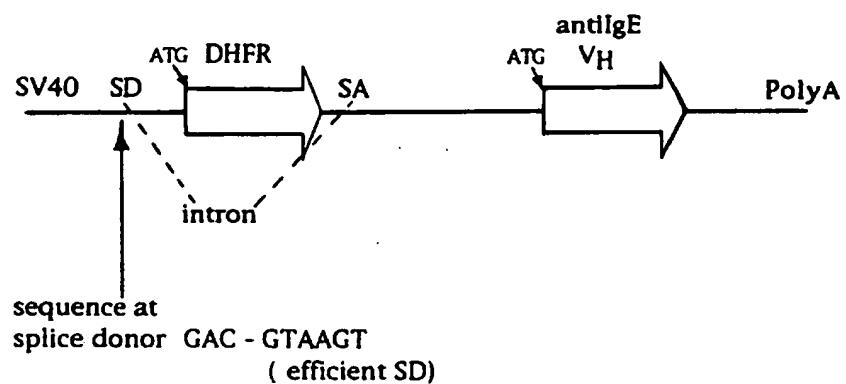
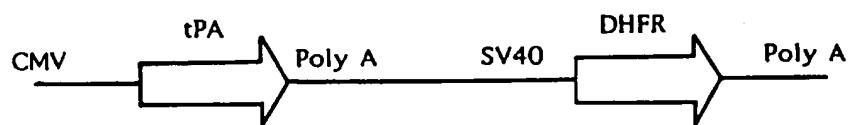
FIG. 1D**FIG. 2**

FIG. 3A

aluI
 sstI
 sacI
 hgiJII
 hgiAI/asPHI
 ecI136II
 bsp1286
 bsiHKAII
 bmyI
 banII
 taqI
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 AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCAAATATTA TCATTAGTTA ATGCCCACT AATCAAGTAT CGGTATATA CCTCAAGGG CAATGTTATTG
 scrFI
 mvaI
 ecorII
 dsav
 acII
 bgII bstNI
 sau96I
 haeIII/palI acII
 asuI apyI [dcm+]
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 ACCGCCATT ACCGGGGGA CGACTGGCG GGTRGCTGGG GCGGGTAAC TGCAGTTATT ACTGCATAAC AGCGTATCAT TGCGGTTATC CCGAAAAGGT
 maeII
 hinII/acyI
 ahaII/bsaHI
 atIII
 bgII
 asuI apyI [dcm+]
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 scrFI
 mvaI
 ecorII
 acII
 bgII dsav
 sau96I bstNI
 haeIII/palI acII
 asuI apyI [dcm+]
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 TTTACCGGGC GGACCGTAAT ACGGGTCTAG TACTGGAAATA CCCTGAAAGG ATGAGATGC ATGATAGTC ATAATCAAGTA GCGATAATGG TACCACTACG

thaI
 fnuDII/mvnI
 bstUI
 bsh1236I
 acII maeII
 tru9I
 maeI
 nseI
 speI
 aseI/asnI/vspI
 bsII
 maclII
 hinII/acyI
 ahaII/bsaHI
 atIII
 maeII
 rsal
 csp6I
 ndeI
 rsal
 csp6I
 aatII
 maeII
 hinII/acyI
 ahaII/bsaHI
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 maeII
 rsal
 csp6I
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 csp6I
 maeII
 snaBI
 bsaAI
 csp6I
 bsrl nlaiI
 styI
 nlaiI

FIG. 3B

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rsal	maelI	nlalI
csp6I	hinII/acyI	ahalI/bsaHI
	hinfI	hgICl
	acII	banI
	bsmAI	
	bsmAI	
	acII	
	hgal	
	ccatt	
	aaatgggg	
	tacccgtt	
	ccgtgggg	
	tctataag	
	cagatattc	
	gtctcgagca	
	haellI/palI	
	mcrlI	
	eagI/xmaIII/ecclXI	
	eaelI	
	cfriI	
	fnu4HI	
	aciI	
	thaI	
	fnuDII/mvnI	
	sacII/ssstII	
	nspBII	
	ksplI	
	scrFI	
	dsalI	
	ncrlI	
	bgllI	
	bstUI	
	msplI	
	sau3AI	
	mboI/ndeII(dam-)	
	hpaII	
	dpnI(dam+)	
	dsav	
	dpmI(dam-)	
	bsh1236I	
	alwlI(dam-)	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
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	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
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	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
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	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
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	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
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	bpwAI	
	bbstI	
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	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
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	caullI	
	alwlI	
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	alwlI	
	aciI	
	caullI	
	msplI	
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	caullI	
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	hpaII	
	bpwAI	
	bbstI	
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	alwlI	
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	msplI	
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	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	
	msplI	
	mboII	
	hpaII	
	bpwAI	
	bbstI	
	mnII	
	caullI	
	alwlI	
	aciI	
	caullI	

FIG. 3C

tfiI acII
 thaI hinII fnuDII/mvnI
 bstU I bsvI
 bsh1236I rsal
 csp6I scfI mnII
 701 TTGGAAACGGG GATTCCCCCGT GCCCAAGAGTG CTGTAAGTAC CGGCCATTAGA GCGATAAGAG GATTTTATCC CCGCTGCCAT CAGGGTCCA GCAATGAACT
 AACCTTGGCG CTAAGGGCA CGGTTCTCAC GACATTATG GCGGATATCT CGCTTATCTC CTAAAATAGG GGCACGGTA GTACCAAGGT GGTAACCTGA
 fnu4HI
 nspBII
 acII
 nlaIII
 taqI
 thaI
 fnudII/mvnI
 bstU I
 bsh1236I
 mluI
 afIIII
 rsal
 csp6I
 xbaI
 scfI
 mnII ddeI asp700
 801 GCATCGTCGC CGTAGCCCCAA AAATATGGGA TTGGCAAGAA CGGAGACCTA CCCTGGCCCTC CGCTCAGGAA CGCGTTCAAG TACTCCCAA GAATGACCAC
 CGTAGCAGGG GCACAGGGTT TTATACCCCT AACCGTTCTT GCCTCTGGAT GGGACGGGAG GCGAGTCCTT GCGCAAGTC ATGAAAGTTT CTTACTGGTG
 scfI
 mvaI
 ecORII
 dsAV
 bstN I
 apyI(dcm+)
 901 AACCTCTTCGA GTGGAAAGGTAA AACAGAAATCT GGTGATTATG GTAGGAAAAA CCTGCTTCCTGAG AAGAATCGAC CTTAAAGGA CAGAMTAAT
 TTGGAGAAGT CACCTTCAT TTGTCTTAGA CCACTAATAC CCATCCTTT GGACCAAGAG GAAATTCT GTCTTAATTA
 tfiI
 hinII
 alwNI
 hphI
 eco57I
 mboII
 earI/ksp632I
 mnII
 1001 ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTCTTG TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CGGGAATTGG
 TATCAAGAGT CATCTCTGA GTTTCTTGT GGTCGCTCTC GAGTAAGAG AACGTTCTCA AACCTACTAC GGAAATTCTGA ATAACCTGTT GGCCCTTAACC
 fnu4HI
 nspBII
 acII
 nlaIII
 taqI
 thaI
 fnudII/mvnI
 bstU I
 bsh1236I
 mluI
 afIIII
 rsal
 csp6I
 xbaI
 scfI
 mnII ddeI asp700
 scfI
 mvaI
 ecORII
 dsAV
 bstN I
 apyI(dcm+)
 tfiI
 hinII
 sexAI
 ddeI
 hinfI
 mboII
 taqI
 mspI
 msel
 ahaiII/draI
 aseI/asnI/vspI
 CTTAAAGGA CAGAMTAAT
 tfiI
 tru9I
 mspI
 msel
 ahaiII/draI
 aseI/asnI/vspI
 CTTAAAGGA CAGAMTAAT
 aluI
 sstI
 sacI
 hgiJII
 hgiAI/asphi
 ec1136II
 bsp1286
 bsiHKAI
 bmyI
 banII
 mnII
 bstX I
 fokI
 afII/bfri
 1001 ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTCTTG TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CGGGAATTGG
 TATCAAGAGT CATCTCTGA GTTTCTTGT GGTCGCTCTC GAGTAAGAG AACGTTCTCA AACCTACTAC GGAAATTCTGA ATAACCTGTT GGCCCTTAACC

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FIG. 3D

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FIG. 3E

SUBSTITUTE SHEET (RULE 26)

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FIG. 3E

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FIG. 3G

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FIG. 3H

2601 mspI hpaII
bsII scrFI nciI nlaiV
dsAV acII cauII bsrBI bsII acII aluI tfII aluI
GAGAGGGTT CCTGCTCCG GCGCATACCA TCAGCTCCTG CTGGATTCTC TCTGCCGCC ACTGCTTCCA
CTCTCGCAA GGACACGCC CGCTAGGGC GTTACGAAAG AGACGGGG TGACGAAGGT

scrFI mvaI
ecoriI dsAV bstNI apyI [dcm+]

sau3AI mboI / ndelli [dam-] dpnI [dam-] dpnII [dam-]
hphI [dam-] hphI [dam-] cauII asuI dsAV avail mnII apoI tagI
ACCTGACGGT GATCTGGGC AGAACATACC GGGTGGTCCT GGAGGAAT TTGAGTGA AAAATACTT
CTAGAACCCG TCTTGATGG CCCACAGGG ACCGCTCTC CTGCTCTTA AACTTCAGCT TTTATGTA

scrFI pstI fnu4HI bbvI pvuII fru4HI fru4HI
mvaI ecoRII dsAV bstNI fspBII bsaJI bbvI
apoi tagI hincII bsgI aluI hincII hincII apyI [dcm+]
GTCCTAAGG AATTGGATGA TGACACTTAC GACAATGACA TTGGATCTGT GCAGCTGAAA TCGGATCTGT CCGCTCTGC CCAGGAGAGC AGCGTGGTCC
CAGGTATTCC TTAGCTACT AGCTGAAAT CTGTCGACTTT AGCCTAAGCA AACGGGACGA CGTCGACTT AACGGGACAG AGCTTAAGCA GGGGACACG GGTCCTCTCG TCGCACCCG

FIG. 31

FIG. 3I		SUBSTITUTE SHEET (DRAFT 2)											
aluI													
sstI													
sacI													
hgjII													
mspI	scfI	pvuII											
hpall	pstI	fnu4HI											
scrFI	bsPMI	bbVI											
ncII	sau96I	nsPBII											
dsAV	avaII	fnu4HI	bsRI										
cauII	asul	bbVI	mspI										
draIII	bsII	bsGI	aluI	hpall									
2901	GCACTGTGTC	CCTTCCCCCG	GCGGACCTGC	AGCTGCCCGA	CTGGACGGAG	TGTGAGCTCT	CCGGCTACGG	CAAGCATGAG	GCCTTGTC	CYTCTATTC	CCGACACAT	GGAAACAGAG	GAAGATAAG
CGTGACACAC	CGCTGGACG	CGAAGGGGGC	CGCTGGAC	TGACGGCCCT	GACCTGCCTC	ACACTCGAGA							
mspI	scfI	pvuII											
hpall	pstI	fnu4HI											
scrFI	bsPMI	bbVI											
ncII	sau96I	nsPBII											
dsAV	avaII	fnu4HI	bsRI										
cauII	asul	bbVI	mspI										
draIII	bsII	bsGI	aluI	hpall									
3001	GAGGGGCTG	AAGGAGGCTC	ATGTCAGACT	GTACCCATTC	AGCCGCTGCA	CATCACAA	TTTACTAAC	AGAACAGTCA	CCGACACAT	GGTGTGTCT	CCGATGTC	GGTGTGTCT	GGTGTGTCT
CTCTGGCGAC	TTCCTCCGAG	TACAGTCTGA	CATGGGTAGG	TCGGGGACGT	TGAGTGTGT	AAATGAAATG	TCTGTGTGT	GTAGTGTGT	AAATGAAATG	TCTGTGTGT	GGTGTGTCT	GGTGTGTCT	GGTGTGTCT
mspI	scfI	pvuII											
hpall	pstI	fnu4HI											
scrFI	bsPMI	bbVI											
ncII	sau96I	nsPBII											
dsAV	avaII	fnu4HI	bsRI										
cauII	asul	bbVI	mspI										
draIII	bsII	bsGI	aluI	hpall									
3101	GGAGACACTC	GGAGGGCGG	GGCCCGAGCA	AACCTGGCAGC	AACCTGGCAGC	ACGCTGTC	GGGGATTG	GGAGGGCCCC	TGCTGTCT	GAACGATGCC	CGATGACTT	CTTGCTACCG	GGCTGACTGA
CCCTGTGAG	CCTCGCCGCC	CGGGTCCGT	TGAACTGTC	TGCGGACGGT	TGCGGACGGT	CCCGCTTAAGC	CCCGCTTAAGC	CCCGCTTAAGC	CCCGCTTAAGC	CCACACAGA	CCACACAGA	CCACACAGA	CCACACAGA

FIG. 3J

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FIG. 3K

styI
 acI
 fnu4HI sau96I
 bglI nlaIV
 sfi I ncoI haellI/palI
 haellI/palI
 eaeII dsalI asul
 cfrI bsalI
 3601 GATGGCGCC ATGCCAAC TTGTTTGT CAGCTTATAA TAAAGCAATAA TGTTTACAAA GCATCACAAA AAAGCATT TTTCAGTCGA
 CTACCGGGG TACCGGGTAC ACAAATAAC GTCGAATT ACCAATGTT ATTTCGTT CGTAGTGTAA AGAGTGACT
 sau3AI
 mboI/ndeII [dam-]
 dpnI [dam+]
 dpnII [dam-]
 pvuI/bspCI
 mcrI
 taqI [dam-] tru9I
 clal/bsp106 [dam-]
 sau3AI msel
 mboI/ndeII [dam-]
 dpnI [dam+] xbaII
 dpnII [dam-] asel/asnl/vspI
 rnaI
 maeI
 3701 TTCTAGTTGT GGTTGTCCA AACTCATCAA TGATCTTAT CATGTCTGGA TCGATCGGA ATTAATTGG CGCAGCACCA TGGCCTGAA TAACCTCTGA
 AAGATCAAACA CCAAACAGGT TTGAGTAGTT ACATAGAATA GTACAGACCT AGCTAGCCCT TATTAAGCC GCGTCGTGGT ACCGGACTTT ATTGGAGACT
 rsal
 csp6I
 nlaIV
 kpnl
 hgiCI
 banI
 asp718
 acc65I
 mnII
 3801 AGAGGAACT TGGTTAGGTA CCTTCTGGA CGGAGAGAC CAGCTCTGGA ATGTTGTCA GTTACGGGTGTT GGAAAGTCC
 TTCTCTGGA ACCAATCCAT GGAAGACTCC GCCTTCTGGA TACACAGT CATTACACCT CATTACACAGT CATTACACAGT CATTACACAGT
 aluI
 pvuII
 rsPBII
 apyI [dcm+]
 bsalI
 nlaIV
 scrFI
 mvaI
 ecoRII
 dsalV
 bstNI
 apyI [dcm+]
 bsalI
 mnII
 ddeI acII
 rsPBII
 mnII
 3901 AGAGGAACT TGGTTAGGTA CCTTCTGGA CGGAGAGAC CAGCTCTGGA ATGTTGTCA GTTACGGGTGTT GGAAAGTCC
 TTCTCTGGA ACCAATCCAT GGAAGACTCC GCCTTCTGGA TACACAGT CATTACACCT CATTACACAGT CATTACACAGT CATTACACAGT

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FIG. 3

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FIG. 3M

hinPI
hhAI/cfoI
nlarIV
nari
kasi
hinII/acyI
hgICl
haeII acII
bani sfANI
ahaiI/bsaHI
sfANI acII
GGAAATGGCG CCTGATGCC TATTTCCTCC TTACGCATCT GTGCCTATT

4301 CGAAGAGGCC CGCACCGATC GCCCCCTCCCA ACAGTTGGT AGCCTGATG CGCAACTTAC CGCTTACGCC ATAAAAGGG AATGCCTAGA CACGCCATAA
 earI/ksp632I mcrI pVU/bSPCI bgII acII
 GCTTCTCCGG GCGTGGCTAG CGGGAAAGGT

hinPI
thaI
fnUDII/mvnI bstUI scfI
bsh1236I hhaI/cfoI
rsal hhaI/cfoI
csp6I bsII acII
 acII maeII
 TCACACCGCA TACGTCAAAG CAACCATAGT ACCGGCCCTG TAGCGGGCGCA TAAAGCCGGG CGGGTGTGGT GTTACCGCC AGCGTGACCG CTACACTGGC
 AGTGTGGCGT ATGCACTTTC GTGGGTATCA TGGGGTACCA ATCGCCCGGT AATTGCGCC GCCCACACCA CAAATGCGCC TCGCACGGC GATGTGAACG

hinPI
hhAI/cfoI
 rmaI
 hinPI haeII
hhAI/cfoI bsrBI
 haelI maeII acII
 CAGGCCCTA GGCGCCCTA CTTTCGCTT CTCCCTTCC TTTCTCGCCA CGTTTCGCGG CTTTCCCGT CAAGCTCTAA ATCGGGGGET CCCTTTAGGG

GTCGGGGAT CGGGGGAT CCCAAGAAC TTGATTTGG TGATGGTCA CGTAGCTGGGC CATGCCCTG ATAGACGTT TTGCCCCGA GGGAAATCCC

nlarIV
hgICl tagI
bani mnII
 4601 TTCCGATTTA GTGGCTTACG GCACCTCGAC CCCAAGAAC TTGATTTGG TGATGGTCA CGTAGCTGGGC CATGCCCTG ATAGACGTT TTGCCCCGA
 AAGGCTAAAT CACGAAATGC CGTGGAGCTG GGTTTTTG ACTAAACCC ACTACAAAGT GCATCACCCG TATCTGCCA AAAGGGGAA

maeII pleI
 drdI hinFI maeII
 4701 TGACGGTGGAA GTCCACGGTC TTAATAGTG GACTCTTTT CCAAACACTGGA ACAACACTCA ACCCTATCTC GGGCTATTCT TTGATTATAGGTTT
 ACTGCAACCT CAGGTGCAAG AAATTATCAC CTGAGACAA GGTTTGACCT TGGTGTGAGT TGGTGTGAG CCCGATAAGA AACCTAAATA TTCCCTAAAA

pleI
 maeII
 hinFI
 bsRI
 bsII
 avai

hinPI
hhAI/cfoI
thaI
fnUDII/mvnI bstUI
bsh1236I hhaI/cfoI
tru9I acII
mseI bsh1236I acII
 maeIII bbyI maeIII
 maelI bbyI maeIII
 mspl
hpII
 naeI
 maeII cfr10I
 aluI
 banII
 banII
 nlarIV
 hgICl
 bsp1286
 bmyI
 banII
 nlarIV
 haellI/palI
 sau96I
 asuI
 hphI
 draII
 bsaAI
 maelI
 haellI/palI
 tru9I
 pleI
 maeII
 drdI
 4701

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FIG. 3Z

FIG. 30

nlaIV
 acII
 thaI
 fnuDI I/mvni I
 bstUI
 bsh123 6 I
 hinPI
 hhaI/cfoI
 mbII I
 ssPI
 5201 CTTTTCGGGG AAATGTTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTGCC TTATTCCCTT TTTGCCCA TTTGCCCTC CTGTTTTC TCACCCAGAA
 TATTATACT TTTCTCTT CATACTCATTA AGTTGTAAG GCACAGGGG AATAAGGAA AAAACCGGT AAAACAAAG AGTGGTCTTCA
 hgiAI/asPHI
 bsp1286
 sau3AI
 bsiHKAI
 mbII/I ndeII [dam-]
 dprI [dam+] bmyI
 dprII [dam-]
 eco57I
 sfaNI
 hphi
 5401 ACGCTGGTGA AAGTAANAGA TGCTGAAAGAT CAGTTGGTG CACGAGTGGG TTACATGAA CTGGATCTCA ACAGGGTAA GATCCTGAG AGTTTCGCC
 TGGGACCACT TTCATTTCT ACGACTCTCA GTGTCACCC ACAGTCACCC ATGTTAGCTT GACCTAGCTT CTAGGAACTC TCAAAAGGG
 rcal
 bspHI
 bsrBI
 bsmAI
 acII nlaIII
 fnu4HI
 acII
 hphi
 5301 ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTGCC TTATTCCCTT TTTGCCCA TTTGCCCTC CTGTTTTC TCACCCAGAA
 TATTATACT TTTCTCTT CATACTCATTA AGTTGTAAG GCACAGGGG AATAAGGAA AAAACCGGT AAAACAAAG AGTGGTCTTCA
 hgiAI/asPHI
 bsp1286
 sau3AI
 rspBII
 mboII/I ndeII [dam-]
 dprI [dam+]
 dprII [dam-]
 bsrI
 dprII [dam-]
 alw44I/snoI
 alw44I [dam-]
 eco57I
 sfaNI
 hphi
 5501 CCGAAGAACG TTTCCAATG ATGAGGACTT TTAAAGTCT GCTATGTTGC GGGTATTAT CCCGTATGAA CGATACACCG CGCCATATA
 GGCTTCTTCG AAAAGTTAC TACTCCGTGA AATTCAGA CGATACACCG CGCCATATA GGCCACTACT GCGCCCCGTT CTGGTTGAGC CAGGGGTTA
 scrfI
 acII
 thaI
 fnuDII/mvni I
 bsh1236 I
 hphi
 hgiI acyI
 hgiI cauli
 ahaII/bsaHI
 bciI
 acII
 rsal
 csp6I
 bsrI
 ddeI
 scaI
 hphi
 maelli
 fokI
 nlaIII
 fokI
 nlaIII
 fnu4HI
 bbVI
 nlaIII
 5601 ACACATTCT CAGAATGACT TGGTTGAGTA CTCACCAAGTC ACAGAAAAGC ATCTTACGGA TGGCATGACA GAAAGAGAAAT TATGCAAGTGC TGCATAACC
 TGTGATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTCAG TAGAATGCTT ACCGTACTGT CATTCCTCTTA ATACGTCAAG ACGGTATTGG

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FIG. 3P

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FIG. 30

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FIG. 3R

FIG. 3S

thaI
 fnuDII/mvnl
 bstUI
 bsh1236I
 hinPI
 hhaI/cfoI
 thaI
 fnuDII/mvnl
 bstUI haellI/pallI aluI
 bsh1236I tru9I pvuI
 mboII hhaI/cfoI mnII eaeI tfI I asel/asnl/vspI
 earI/ksp632I acII cfRI hinFI mseI nspBII
 mnII acII haellI acII cfrI hinfI mseI nspBII
 7101 GTCAGTGAGC GAGGAAGCGG AAGAGGCCCC AATACGAAA CGGCCCTCTCC CGGGGTTC GCGGATTICAT TAATCCAGCT GGCACGACAG GTTCCCGAC CAGTCACTCG CTCCCTCGCC TTATGCGTT GGCGGAGGG GGCGGCAAC CGGGTAACTGA ATTAGGTGCA CGGTGCTGTC CAAAGGGCTG
 scrFI
 mvaI
 ecORII
 dseV
 tru9I
 mseI
 maeIII
 mnII
 hgiCI aspI [dcm]
 nlaIV bstNI
 hpaII
 bani bsaji
 mspl
 acII
 hhaI/cfoI asel/asnl/vspI mnII
 7201 TGGAAAGCGG GCAGTGAGCG CAACGGCAATT ATGTGAGTT ACCTCACTCA TTAGGACCCC CAGGCTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTC ACCTTCGCC CGTCACTCGC GTTGCCTAA TTACACTCAA TGAGGTGAGT AATCCGGGG GTCCGAAATG TGAATAATCGA AGGGCGAGCA TACAACACAC
 tru9I
 mseI
 asel/asnl/vspI
 xmnI
 acII
 bsrBI
 aluI
 nlaIII
 asp700
 7301 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT TAGGAATTAA CTTAACACTC GCCTATTGTT AAAGTGTGTC CTTGTGTCGAT ACTGGTACTA ATGCTTAATT
 >length: 7360

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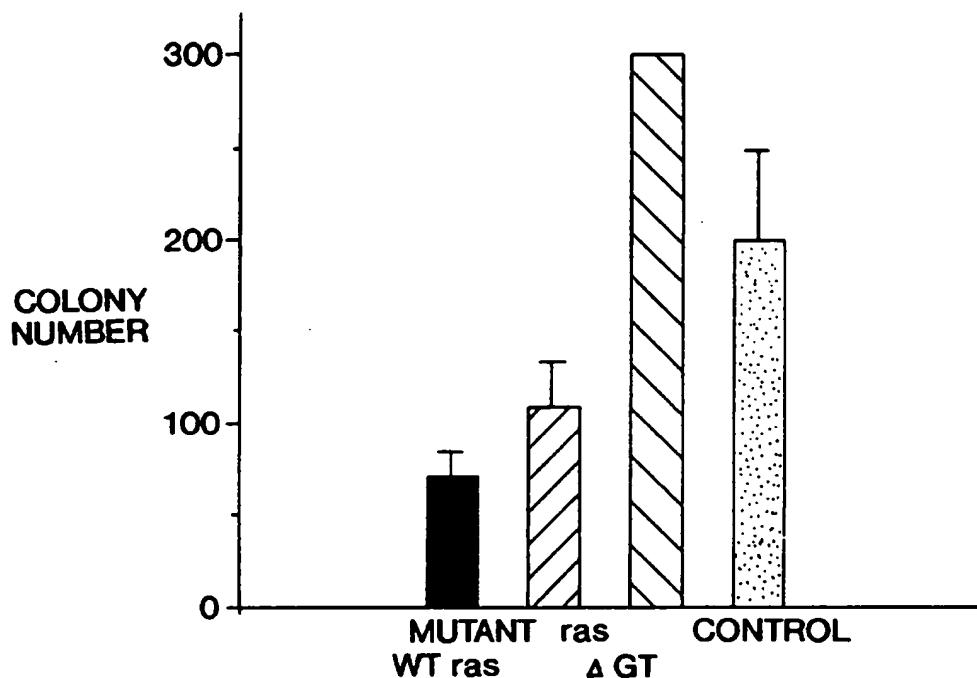


FIG. 4

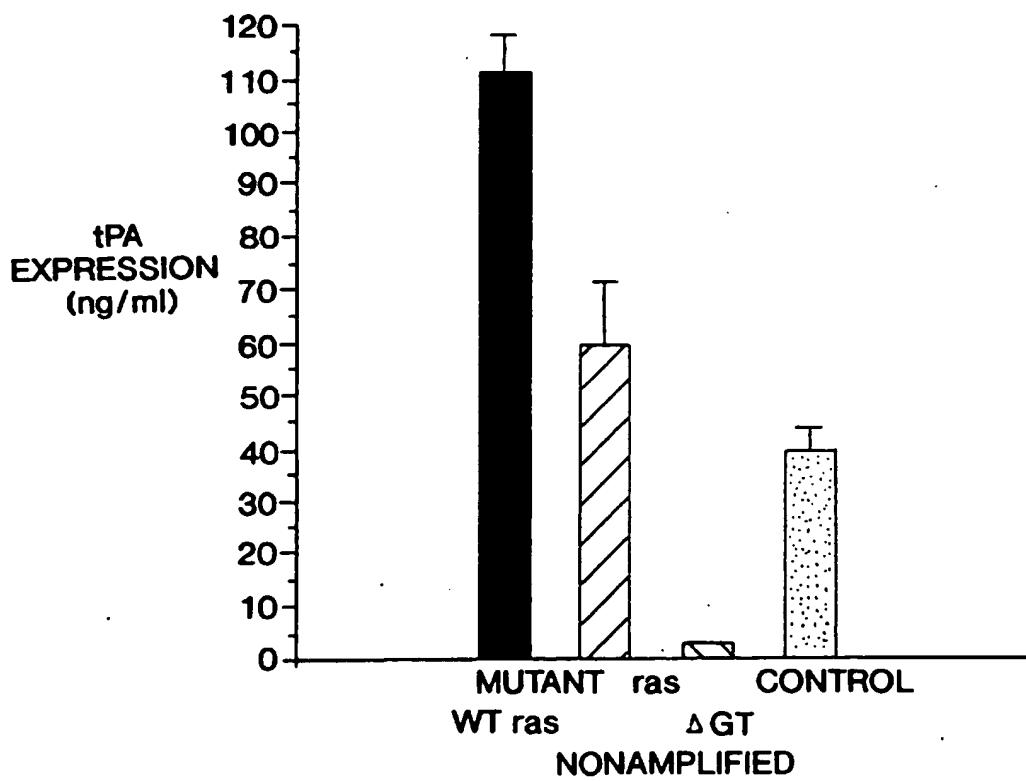
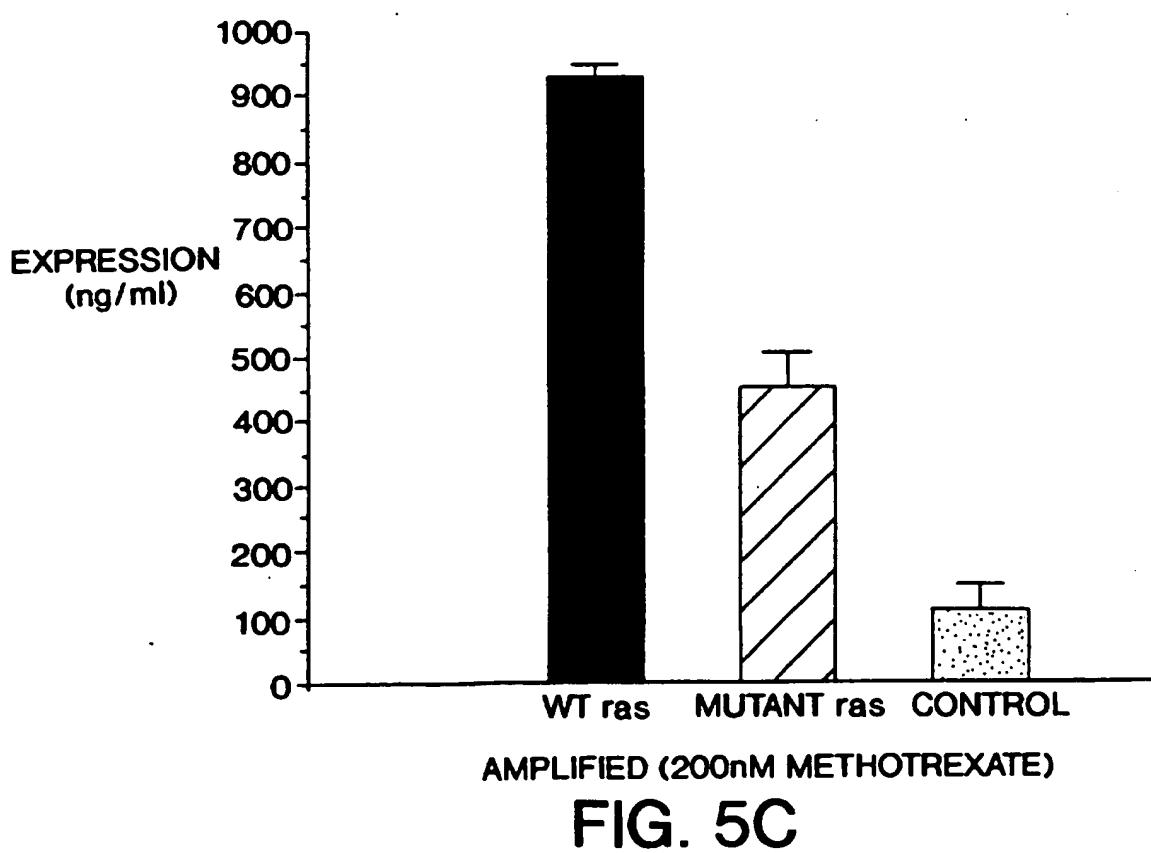
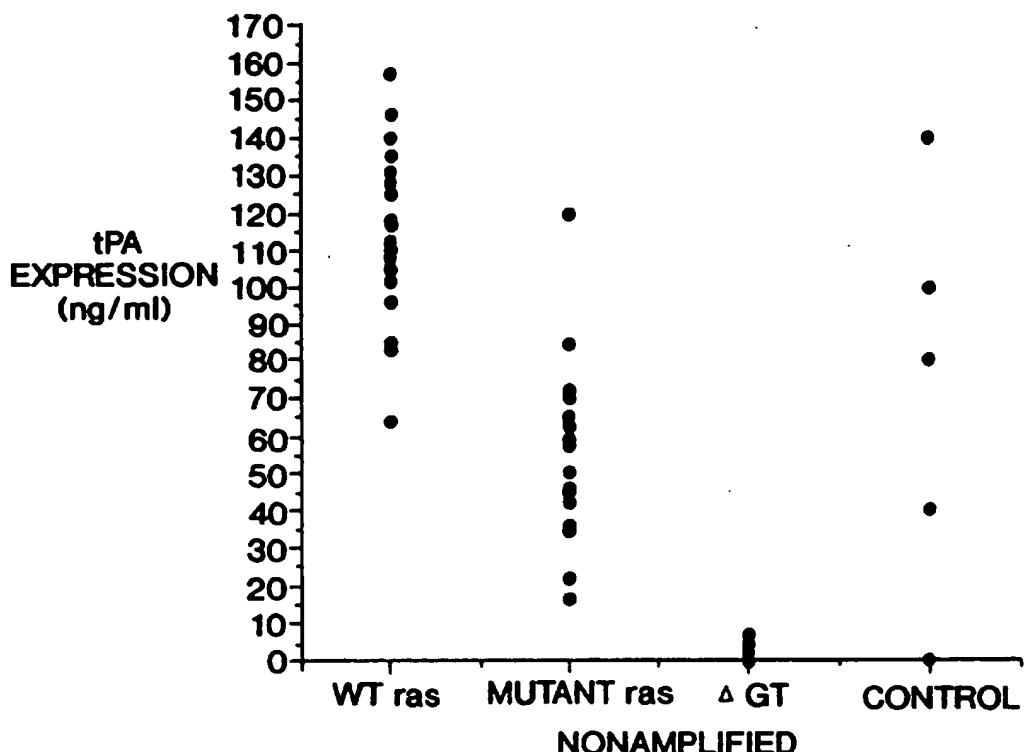


FIG. 5A

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FIG. 6A

aluI
sstI
sacI
hgjII
hgjAI/asPHI
ec1136II
bsp1286
bsiHKAI
bmyI
banII
tagI

1 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATAAT AGTAATCAAT TACGGGGTCA TTAGTCATA GCCCATATAAT GGAGITCCGC GTTACATAAC AAGCTGAGC GGGCTGAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCACTG AATCAAGTAT CGGGTATATA CCTCAAGGGG CAAATGATTG

scrFI

rmaI
tru9I
maeI
mseI
spel
asel/asnI/vspI

101 TTACGGTAAA TGGCCGGCTT AATGGCATT ACCGGCGGA ACCTGAGCTTACCTCA TAAATGCCAT TTGACGGGTG AACCCTCATG TAGTTCACAT AGTACGGTATT ACTGCATACA AGGGTATCAT TCGGGTATAC CCTGAAAGGT

maeII
hinII/acyI
sa96I
haeIII/palI
aciI
asuI apyI [dcm+]
ggctgaccgc
ccaaacgacc
cggcccattg
ccgtcaataa
tgacgtatgt
tcccatatgt
acgccaatag
ggactttcca

201 TTGACGTCAA TGGGTGGAGT ATTACGGTA AACTACGGTA AACTGCCCAC TTGGCAGTAC ATCAAGTGT TCATATGCCA AGTACGGT CTATTGACGT CAATGACGGT AACTGAGTT ACCACCTCA

scrFI
mvaI
ecorII
aciI
bgII bstNI
sa96I
haeIII/palI
aciI
rsal
bgII
asuI apyI [dcm+]
ggctgctggc
ccgactggcc
ccggggcgaa
tttacggat
aatggccattt
tttacggat
aatggccattt

301 AAATGGCCCG CCTGGCCATTA TGCCCGATAG TACTGGCAG TACATCTACG TATTAGTCTACG CCCTTATACG ATGTTATACC ATGTTGATCC TTACCGGGC GGACCGTAAAT ACGGGGTCTAG TACTGGATAATA CCTCGAAAGG ATGAAACGGC ATGTTAGATGC ATAATCAGTA GCGATAATGG TACCACTACG

thaI
fnuDII/mvnI
bstUI
bsh1236I
aciI maeIII

bslI
aciI

maeII
hinII/acyI
ahaiII/bsaHI
atiII
maeIII
maeII
rsal
csp6I
ndel
rsal
csp6I
atiII
maeII
rsal
ndel
rsal
bsaHI
bsaAI
bsaAI
bsrI nlairI
styI
maeII
snabI
csp6I
bsrI nlairI
maeII
rsal
csp6I
bsrI nlairI
ncoI
dsal
phI acI
bsaJI sfanI

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FIG. 6B

FIG. 6C

FIG. 6C

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FIG. 6D

haeIII/pall

scrFI	scrFI	nlaiII	sau3AI
mvaI	mvaI		
ecoriI	ecoriI		
dsAV	tfII	dsAV	pleI
bstNI	nlaiII	bstNI	ddeI
apyI [dcm+]	hinFI	apyI [dcm+]	dpnII [dam-]
GACATAGT AGACATGGTT TGGATAGTCG mnII	GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCCTGTGA CAAGGATCAT	hinFI	maelII alwI [dam-]
GTTCAATTCA TCTGTACCAA ACCTATCAGC CTCGTCAGG ACAATGGTC CTTCGGTACT TAGTGGTCC GTGGAATCT GAGAAACATC GRTCTCTAGTA			
mnII	hinII/acyI	scrFI	
	ahalII/bsaHI	mvaI	
	scrFI	ecoriI	
	mvaI	ecoNI	dsAV
	ecoriI	sau96I	
	dsAV	availI	
	bstNI	bsII	asul mnII
	apyI [dcm+]	bsaJI	bstNI
	maelII	hgaI	ddeI apyI [dcm+]
apoI	afIIII	mnII	cccaggatcc CCAGGGTCC TCTCTGAGGT CCAGGAGGAA
GCAGGAATTG AAAGTGACA GTTTTCCC AGAAATTGAT TTGGGAAAT mnII	ATAAACCTCT CCCAGAAATAC TCTACGAAAG TCTACGAAAG TTTCGATACG TAAATATT		
CGCCCTAAA CTTTCACTGT GCAAAAAGG TCTTTAACTA AACCCCTTTA TATTGGAGA GGTCCTTATG GGAGACTCCA GGTCCTCC			
ppul0I			
	nsII/availII		
sfanI	accI	mbolI	mnII
			alwI
1301 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAG ACTAACAGGA AGATGCTTC AAGTCTCTG CTCCCCCTCT AAAGCTATGC ATTCTTATAA	TCTCTCTTC TGATGTCCT		
TTTCGTTAGT TCATATTCAA ACTTCAGATG	TCTACGAAAG TTCAAGAGAC GAGGGAGGA TTTCGATACG TAAATATT		
fnu4HI			
nlaiII			
styI			
ncI			
dsI			
bsaJI	styI	fnu4HI	
	bsaJI	thaI	
		fnu4II/mvnI tru9I	
		bstUI	
		bsb1236I aseI/asnI/vspI	
		mseI	
1401 GACCATGGGA CTTTGCTGG CTTAGACCC CCTGGCTTC GTAGAACGC GGCTACAATT AATACATAAC CTATGTTAC ATACACATAG ATTAGGTGA	maelII		
CTGGTACCT GAAAACGACC GAAATCTGGG GGAAACGGAAG CAATCTGGC CGATGTTAA TTATGTTAG GAATACATAG TATGTTACG TAAATCCACT	hphI		

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FIG. 6E

haeIII/pali
eael

scrFI	
mvai	
ecoriI	
dsav	
bstNI	
apyI [dcm+]	mnII
gsuI/bpmI	bsaJI
scfI	mnII
fokI	hincII/hindII
1501 CACTATAGAA TAACATCCAC TTTGCCTTC TGTCACAGG AGAGGAAAG AACGGTGTCC ATAGTAGGT CCAGTGAGG TGGAGCCAAG ATAGCTAACT TAAGGGCCC GTATCGACAG	clal/bspI06 bsaJI aluI
gsuI/bpmI	taqI apoI cauII
scrFI	ecoriI dsav
mvai	ecoriI dsaV
mnII	mnII
sau96I	nsPBII
haeIII/pali	hgiAI/asphi
asur	fnu4HI acII
nlaiII	fnu4HI bspI1286
1601 TGGCATGGC CTCTCCACCG TGCTGACT GCTGTGCC CGTAGCTGGT GTGAGCTGG ACTGGTCCCT ACCGTACCCG GAGGTGGC ACGGACTGA GGACGACGG ACCTCGACAR CCACCCATT ATGGGAGTC CCCATTAAC TGACCAAGGA	bbVI fnu4HI bsiHKAI aluI bmyI apyI [dcm+]
rmaI	bsu36I/mstII/sauI
maeI	bsu36I/mstII/sauI
styI	bsu36I/mstII/sauI
bsaJI	bsu36I/mstII/sauI
blnI	bsu36I/mstII/sauI
avrI	bsu36I/mstII/sauI
1701 CACCTAGGG ACAGGGAGAA GAGAGATAAT GTGTGTCCCT AGGAAATAATA TATCCACCT CAAATAATT GATTGTCTG TACCAATGTC CACAAGGAA	earI/ksp632I taqI
GTGGATCCCC TGTCCCTCTT CTCTCTATCA CACACAGGG TTCCCTTTAT ATAGGTGGG GTTTTATAA CCTAAACGAC ATGGTTAACG GTGTTTCCTT	rsal csp6I nlari

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FIG. 6F

FIG. 6G

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6H
E.G.
U

FIG. 61

FIG. 6J

rmaI
 mn1I
 sau3AI
 mboI/ndeII [dam-]
 dpnI [dam+]
 dpnII [dam-] taqI
 alwI [dam-] salI scfI
 nlaIV maeI hinCII/hindII
 bstYII/xbaII accI pstI
 barWI xbaI pleI bsgI
 alwI [dam-] hinFI bspMI
 hindIII bgI I nlaIII
 3101 GGGGATCCCTC TAGAGTCGAC CTGCAGAAAGC TTGCGCCGCCA TGGCCCCACT TGTTTATTGC AGCTTATAAT GGTTACAAT AAAGCAATAG CATCACAAAT
 CCCCTAGGAG ATCTCAGCTG GACGTCTTG AACCGGGCT ACCGGGTGA ACAATAACG TCGAATATTAA CCAATGTTA TTTCGTTTC GTAGTGTAA

styI
 acI
 fnu4HI sau96I
 sfII ncoI haeIII/palI
 haeIII/palI
 eaeI dsal asuI
 aluI fru4HI
 bbVI
 maeIII
 sau3AI
 mboI/ndeII [dam-]
 dpnI [dam+]
 dpnII [dam-]
 pvuI/bspCI
 mcRI
 taqI [dam-] tru9I
 clai/bspI06 [dam-]
 sau3AI mseI
 mboI/ndeII [dam-]
 dpnI [dam+] xbaII
 dpnII [dam-] asel/asnl/vspI
 sau3AI mseI
 mboI/ndeII [dam-]
 dpnI [dam-] asp700 hhAI/cfoI
 nlaIII alwI [dam-] asp700 hhAI/cfoI
 hinPI

rmaI
 bsmI maeI
 3201 TTACACAAATA AAGGATTTTTT TTACAGTCAT TCTAGTTGGTGGTGGCAT ACTICATCAAT GTATCTTATC ATGTCGGAT CGATCGGGAA TTAATTGGC
 ACTCTTAT TTCTGAAAAA AAGTGACGTA AGATCAACAC CAAACAGGTT TGTAGTAGTTA CATAGAAATAG TACAGACCTA GCTAGCCCTT AATTAAGGCC

rsal
 csp6I
 nlaIV
 kpnI
 hgiCI
 banI
 asp718 mn1I
 mn1I ddeI accI
 acc65I
 nspBII

haeI
 styI
 ncoI
 dsal haeIII/palI
 fnu4HI nlaIII
 bbVI bsaJI mn1I
 3301 GCAGCACCCT GGCTGAAAT AACCTCTGAA AGAGGAACTT GGTTAGGTAC CTCCTGAGGA CCAATCCATG GAAGACTCCG CCTTCTTCTGG TCAGACACCTT ACACACAGTC AATCCCAC

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FIG. 6K

FIG. 6K
 nlaIV
 scrFI
 mval
 ecoRII
 dsAV
 bstNI
 apyI [dcm+]
 bsAJI
 3401 GAAAGTCCCC AGGGTCCCCA GCAGGGAGAA GATGCAAG CTTTCAGGGG TCCGAGGGT CGTCCGGT CATACTTTC GTACGTAG GATTAATCAG CAACCATAG AATTAGTCG TTAACTAGTCG TCGAGGTCCAC ACCTTTAGG GTCGTCGAGG GTGTCGAGG GTCGTCGTC
 sfaNI
 ppu10I
 nsII/availI
 nlaIII
 sphI
 nsPI
 nsPHI
 3501 AAGTATGCCA AGCATGCCATC TCAATTAGTC AGCAACCATA GTCCCCCCC TAACTCCGCC CATCCGCC CTAACTCCGC CCAGTTCGGC CCATTCTCG
 TTTCATACGTT TCGTAGCTAG AGTTAATCAG TCGTTGGTAT CAGGGGGGG ATTGAGGGGG GATTGAGGGG GGTCAAGGGG GGTAAAGGGC
 nlaIII
 styI
 ncoI
 dsAI
 bsAJI
 3601 CCCCATGGCT GACTAATTCTT TTTTATTAT GGAGGGCC AGGGCCGCCTC GGCCCTCTGAG CTATCCAGA AGTAGTGAGG AGGCTTTTT GGAGGGCTAG
 GGGGTACCGA CTGATTAAAA AAAATAATAATA CGTCTCCGGC TCAGGGGGAG CCGGAGACTC GATAAGGTCT TCATCACTCC
 scrFI
 mval
 ecoRII
 dsAV
 bstNI
 apyI [dcm+]
 bsAJI
 3701 GCTTTCGGAA AAAGCTGTT ACAGCTTGGC ACTGGCCGCTG GTTTCACAAC GTCGTGAATG GAAAAACCTT GGCCTAACCC AACCTAATCG CCTTGAGCA
 CGAAAACGTT TTTCGACAAAT TGACGGCCAG CAAATGTTG CAGCACTGAC CCTTTGGGA CGGCAAATGGC TTGAATTTAGC GGAACGTCGTT

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FIG. 6L

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FIG. 6M

FIG. 6M

		thaI	fnuDII/mvnI			
		tru9I	apoi	tru9I	tru9I	tru9I
	mseI			mseI bstUI	mseI	mseI
4301	GGCTTATTCCTT TTGATTATA AGGGATTGT CGGATTTCGG CCTATGGTT AAAAATGAG CTGATTAAC AAAAATTTAA CGCGAATTTC AACAAAATAT CCGATAAGAA AACTAAATAT TCCCTAAAC GGCTAACCCG GGATAACCA TTTTTACTC GACTAAATG TTTTTAATT GCGCTTAAAA TTGTTTTATA	haeIII/pallI	alui	tru9I mseI	apoI bsh1236I	SSPI
		hgiAI/asphI				
		bsp12886				
		bs11HKAI				
		bmyI ddeI				
		apaL1/snoI	rsal	tru9I	maeIII	maeIII
		alw441/snoI	csp6I	mspI	bsrI	bstUI
4401	TAACTTATGG TGCACTCTCA GTACAATCTG CTCTGTGCC GCATAGTTAA GCCAACTTCG CTATCGCTAC GAGACTACGG CGTATCAATT CGGTGTAGGC GATAGGGATG CACTGACCCA GTACCGAAGC	ATGCCAATG	TTAAATACC ACGTGAGAGT CATGTTAGAC	sfANI	bsAAI tth111I/asPI bbVI	bbVI
		hinPI				
		hhaI/cfol				
		thaI				
		fnuDII/mvnI				
		bstUI				
		nspBII	bsh1236I	dsAV fokI	esp3I	fnu4HI
		aciI	hgaI	drdI	maeIII bsmAI	bbVI
4501	CCCCGACACC CGCCCAACACC CGCTGACGGG CCTGTCTGCT GGTACAGAC GCCTGCATCC GCCTACAGAC AAGTGTGAC CGCTCTCCGG AGCTGCATGT GGGCTGTGG CGGGTGTGG GCGACTGCCG GGGACTGCCG GAACAGACGA GGGCCGTAGG CGAATGTCTG TTCGACACTG CGAGAGCCCC TCAGACTA	aciI	cauII	aciI	alul	alul nlalII
		hinPI				
		hhaI/cfol				
		thaI				
		fnuDII/mvnI				
		bstUI				
		bsh1236I				
		hinPI				
		hhaI/cfol				
		thaI				
		fnuDII/mvnI				
		bstUI mnII				
		bsh1236I				
		hinPI				
		hhaI/cfol				
		thaI				
		haeIII/palI				
		mnII				
		sau96I				
		mbII				
		bpuAI				
		bbsI				
		ec00109I/draII				
		GGCTCTGTAT				
4601	GTCAGAGGTTCACCGGTCA TCACGAAAC GCGGAGGCCA GTATTCTGA AGACGAAGG CATAAGAACT TCTGCTTCCC CGGGCCTCGG CGGGCTCCG CAGTCCTCAA AAGTGGCACT TGTAGGTAA ATATCCAA TAGCTACTA	GGCTCTCAA	AAGTGGCACT	GGGGCTTCCC	GGGGCTCCG	GGGGCTCCG

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FIG. 6N

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FIG. 60

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FIG. 6P

5701 AAGCATTGGT AACTGTCAGA CCAGTTAAC TCATATATAC TTAGATTGA
maeIII TTAGAGTA CTGGTTTAG GGAATTGAC TCAAAGCAA GGTGACTCGC AGTCGGGC ATCTTCTA GTTTCCTAG AGAAACTCTAG GAAAAAAGA

5801 ATTAATCTCAT GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGG TCAGAGCCCC TAGAAAGAT CAAGGATCT TCTTGAGATCT CTTTTTTCT
nlaIII TATAGAGTA CTGGTTTAG GGAATTGAC TCAAAGCAA GGTGACTCGC AGTCGGGC ATCTTCTA GTTTCCTAG AGAAACTCTAG GAAAAAAGA

5901 CGGGCATTTAG ACCGCGAACG TTGGTTTTT GGCGGGAT CCTGGCCACC AACACAAACGG CCTAGTTCTC GATCTTGAG AAAAGCTT CCATGACCG
hinPI fnu4HI acII nspBII acII hpII aluI mspl
hhaI/cfoI bbvI acII nspBII ccgggtggg TTGGTTGCC GGATCAAGAG CTACCAACTC TTTTCCAA GCTAACTGCG
6001 CGGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCACCGCTA CGGTGGAT CCTGGCCACC AACACAAACGG CCTAGTTCTC GATCTTGAG AAAAGCTT CCATGACCG
hinPI rnaI haeIII/palI scfI acII mnII
hhaI/cfoI maeI bsI haeI maeIII ecc57I
6001 TTCAGGAGAG CGAGATACC AAATACTGTC CTCTAGTGT AGCCGTAAGT AGCCGACAC TCGAGATAC GCCTACATAC CTGGCTCTGC
 AAGTCGTCG GCGCTATGG TTATGACAG GAAGATCACA TCGGCTCAA CGGATGTTGA GACATGTTG CGGATGTTG GAGGAGACG

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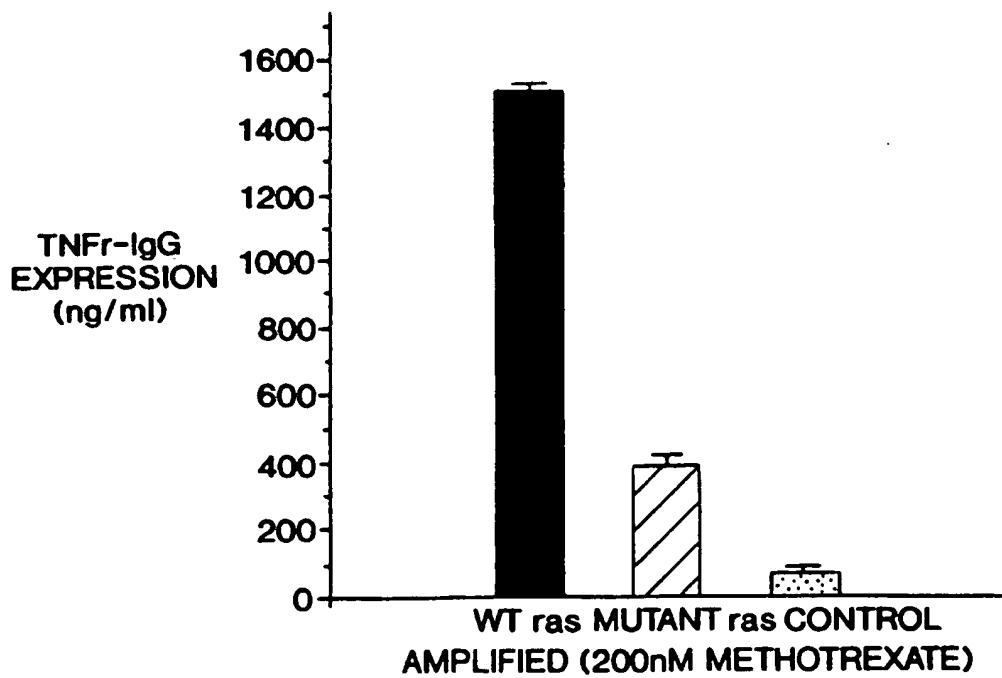
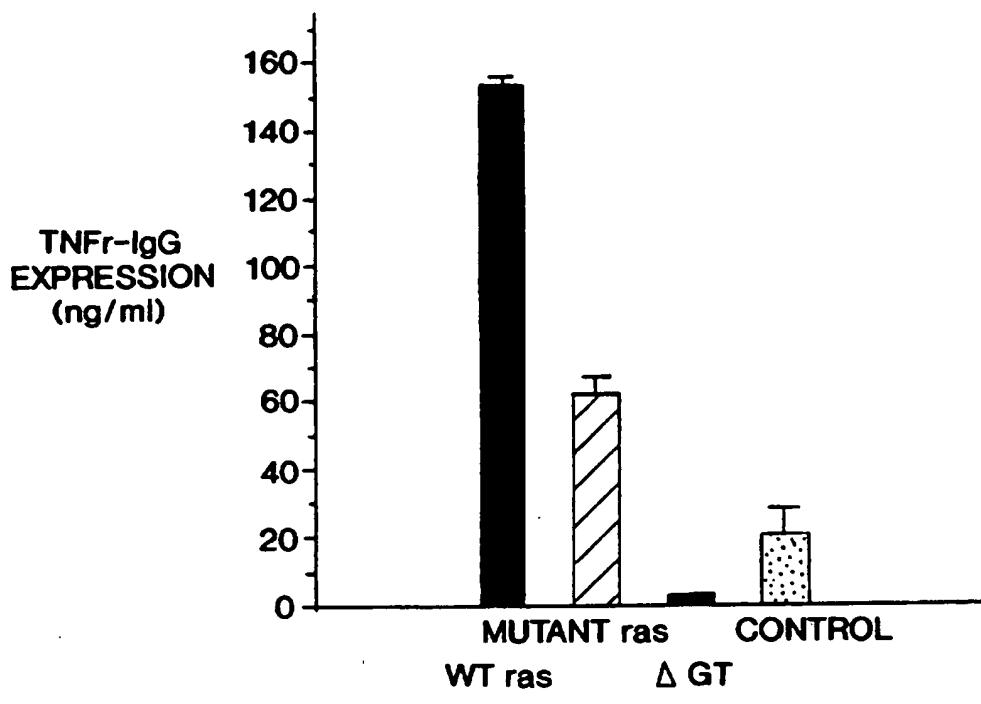
FIG. 60

scrFI	aciI	nlaIV	scrFI	aciI	nlaIV
nciI	nspBII	fnu4HI	nciI	nspBII	fnu4HI
mspI	nsbBI	bbvI	mspI	nsbBI	bbvI
hpall	bspI	bspI	hpall	bspI	bspI
dsav	bspI	bspI	dsav	bspI	bspI
cauII	bspI	bspI	cauII	bspI	bspI
hsfI	bspI	bspI	hsfI	bspI	bspI
maelli	bspI	bspI	maelli	bspI	bspI
maelli	bspI	bspI	maelli	bspI	bspI
6101 TAATCCTGTT ACCAGTGGGT GCTGCCAGTG GCGATAAGTC GTGCTTACG GGGTTGGACT CAAGACGATA TTACCGGAT AAGGCAGC GGTGGGGCTG ATTAGGACAA TGGTACCGA CGACGGTCAC CGCTATTAG CACAGAATGG CCCAACCTGA GTTCTGCTAT CAATGGCTTA TTCCGGTGTG CCAGCCCCGAC	6101 TAATCCTGTT ACCAGTGGGT GCTGCCAGTG GCGATAAGTC GTGCTTACG GGGTTGGACT CAAGACGATA TTACCGGAT AAGGCAGC GGTGGGGCTG ATTAGGACAA TGGTACCGA CGACGGTCAC CGCTATTAG CACAGAATGG CCCAACCTGA GTTCTGCTAT CAATGGCTTA TTCCGGTGTG CCAGCCCCGAC	6101 TAATCCTGTT ACCAGTGGGT GCTGCCAGTG GCGATAAGTC GTGCTTACG GGGTTGGACT CAAGACGATA TTACCGGAT AAGGCAGC GGTGGGGCTG ATTAGGACAA TGGTACCGA CGACGGTCAC CGCTATTAG CACAGAATGG CCCAACCTGA GTTCTGCTAT CAATGGCTTA TTCCGGTGTG CCAGCCCCGAC	6101 TAATCCTGTT ACCAGTGGGT GCTGCCAGTG GCGATAAGTC GTGCTTACG GGGTTGGACT CAAGACGATA TTACCGGAT AAGGCAGC GGTGGGGCTG ATTAGGACAA TGGTACCGA CGACGGTCAC CGCTATTAG CACAGAATGG CCCAACCTGA GTTCTGCTAT CAATGGCTTA TTCCGGTGTG CCAGCCCCGAC	6101 TAATCCTGTT ACCAGTGGGT GCTGCCAGTG GCGATAAGTC GTGCTTACG GGGTTGGACT CAAGACGATA TTACCGGAT AAGGCAGC GGTGGGGCTG ATTAGGACAA TGGTACCGA CGACGGTCAC CGCTATTAG CACAGAATGG CCCAACCTGA GTTCTGCTAT CAATGGCTTA TTCCGGTGTG CCAGCCCCGAC	6101 TAATCCTGTT ACCAGTGGGT GCTGCCAGTG GCGATAAGTC GTGCTTACG GGGTTGGACT CAAGACGATA TTACCGGAT AAGGCAGC GGTGGGGCTG ATTAGGACAA TGGTACCGA CGACGGTCAC CGCTATTAG CACAGAATGG CCCAACCTGA GTTCTGCTAT CAATGGCTTA TTCCGGTGTG CCAGCCCCGAC
hgiAI/asphi	hgiAI/asphi	hgiAI/asphi	hgiAI/asphi	hgiAI/asphi	hgiAI/asphi
bspI1286	bspI1286	bspI1286	bspI1286	bspI1286	bspI1286
bsiHKAI	bsiHKAI	bsiHKAI	bsiHKAI	bsiHKAI	bsiHKAI
bmyI	bmyI	bmyI	bmyI	bmyI	bmyI
apaLI/snoI	apaLI/snoI	apaLI/snoI	apaLI/snoI	apaLI/snoI	apaLI/snoI
a1w44I/snoI	a1w44I/snoI	a1w44I/snoI	a1w44I/snoI	a1w44I/snoI	a1w44I/snoI
aluI	aluI	aluI	aluI	aluI	aluI
ggccgggt	ggccgggt	ggccgggt	ggccgggt	ggccgggt	ggccgggt
tcgtgcacac	tcgtgcacac	tcgtgcacac	tcgtgcacac	tcgtgcacac	tcgtgcacac
agcccccca	agcccccca	agcccccca	agcccccca	agcccccca	agcccccca
tcgggtcgaa	tcgggtcgaa	tcgggtcgaa	tcgggtcgaa	tcgggtcgaa	tcgggtcgaa
tgggtgtgt	tgggtgtgt	tgggtgtgt	tgggtgtgt	tgggtgtgt	tgggtgtgt
6201 AACGGGGGGT TCGTGCACAC AGCCCCCGTT GGAGCGAACG ACTTACACCG CCTACAGGATA CCTACAGGCT GAGCATTGAGT TTGACTCTT GGATGTGGC	6201 AACGGGGGGT TCGTGCACAC AGCCCCCGTT GGAGCGAACG ACTTACACCG CCTACAGGATA CCTACAGGCT GAGCATTGAGT TTGACTCTT GGATGTGGC	6201 AACGGGGGGT TCGTGCACAC AGCCCCCGTT GGAGCGAACG ACTTACACCG CCTACAGGATA CCTACAGGCT GAGCATTGAGT TTGACTCTT GGATGTGGC	6201 AACGGGGGGT TCGTGCACAC AGCCCCCGTT GGAGCGAACG ACTTACACCG CCTACAGGATA CCTACAGGCT GAGCATTGAGT TTGACTCTT GGATGTGGC	6201 AACGGGGGGT TCGTGCACAC AGCCCCCGTT GGAGCGAACG ACTTACACCG CCTACAGGATA CCTACAGGCT GAGCATTGAGT TTGACTCTT GGATGTGGC	6201 AACGGGGGGT TCGTGCACAC AGCCCCCGTT GGAGCGAACG ACTTACACCG CCTACAGGATA CCTACAGGCT GAGCATTGAGT TTGACTCTT GGATGTGGC
mspI	mspI	mspI	mspI	mspI	mspI
hpall	hpall	hpall	hpall	hpall	hpall
bsII	bsII	bsII	bsII	bsII	bsII
fnu4HI	fnu4HI	fnu4HI	fnu4HI	fnu4HI	fnu4HI
aciI	aciI	aciI	aciI	aciI	aciI
6301 GGGAGAAAGG CGGACAGGTAA GCAGGGTCG GACAGGAGA CGGCACGAGG GAGCTTCCAG GGGAAAACGCC CTGGTACCTCT TATAGTCCCT	6301 GGGAGAAAGG CGGACAGGTAA GCAGGGTCG GACAGGAGA CGGCACGAGG GAGCTTCCAG GGGAAAACGCC CTGGTACCTCT TATAGTCCCT	6301 GGGAGAAAGG CGGACAGGTAA GCAGGGTCG GACAGGAGA CGGCACGAGG GAGCTTCCAG GGGAAAACGCC CTGGTACCTCT TATAGTCCCT	6301 GGGAGAAAGG CGGACAGGTAA GCAGGGTCG GACAGGAGA CGGCACGAGG GAGCTTCCAG GGGAAAACGCC CTGGTACCTCT TATAGTCCCT	6301 GGGAGAAAGG CGGACAGGTAA GCAGGGTCG GACAGGAGA CGGCACGAGG GAGCTTCCAG GGGAAAACGCC CTGGTACCTCT TATAGTCCCT	6301 GGGAGAAAGG CGGACAGGTAA GCAGGGTCG GACAGGAGA CGGCACGAGG GAGCTTCCAG GGGAAAACGCC CTGGTACCTCT TATAGTCCCT
ccctcttcc	ccctcttcc	ccctcttcc	ccctcttcc	ccctcttcc	ccctcttcc
gcccattcat	gcccattcat	gcccattcat	gcccattcat	gcccattcat	gcccattcat
ttgtgttgt	ttgtgttgt	ttgtgttgt	ttgtgttgt	ttgtgttgt	ttgtgttgt
taqI	taqI	taqI	taqI	taqI	taqI
nlaIV	nlaIV	nlaIV	nlaIV	nlaIV	nlaIV
aciI	aciI	aciI	aciI	aciI	aciI
fnu4HI	fnu4HI	fnu4HI	fnu4HI	fnu4HI	fnu4HI
bsH1236I	bsH1236I	bsH1236I	bsH1236I	bsH1236I	bsH1236I
aciI	aciI	aciI	aciI	aciI	aciI
thai	thai	thai	thai	thai	thai
bsMII/mvnI	bsMII/mvnI	bsMII/mvnI	bsMII/mvnI	bsMII/mvnI	bsMII/mvnI
bstUI	bstUI	bstUI	bstUI	bstUI	bstUI
bsI	bsI	bsI	bsI	bsI	bsI

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FIG. 6R

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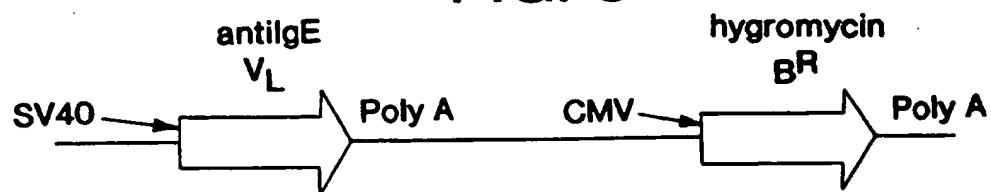
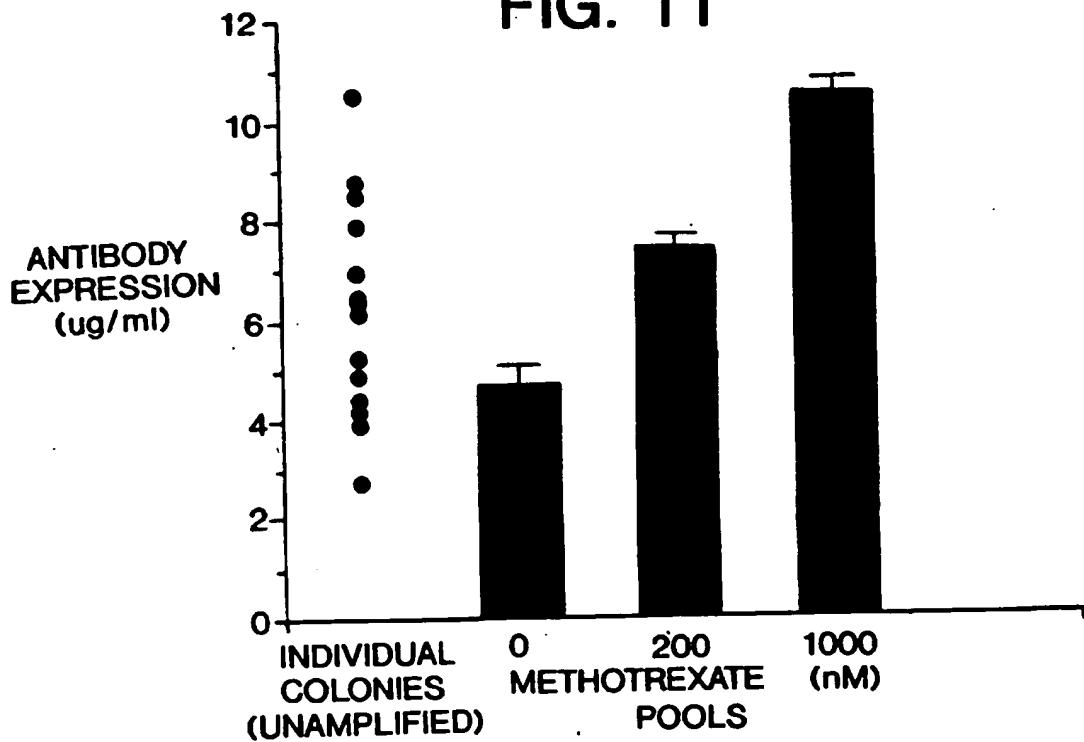
FIG. 8**FIG. 11**

FIG. 9A

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FIG. 9B

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FIG. 9C

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FIG. 9D

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FIG.

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FIG. 9F

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FIG. 9G

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FIG. 9

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FIG. 91

FIG. 9J

2801	AAGGCAATAG CTCACAAATA AAGCATTTT TCTACTCAT TCTAGTGTG TTCCGTAATC GTCAGTGTAA AGCTGTTTAA GTCAGTGTAA AGCTGACTA AGATCAACAC CAAACACCTT TGAGTAGTTA CATAGAAATAG TACAGACCTA	tru9I	nlaIV	rsal
		sau3AI	asel/asnl/vspI	csp6I
		mboI/ndeII [dam-]	styI	nlaIV
		dpnI [dam+]	ncoI	kpnI
		dpnII [dam-]	fnu4HI haelI	hgIC1
		pvuI/bspCI mseI	bbvI dsal haelli/pali	banI
		mcrI asp700	hinPI bsaJ1	asp718
		CGATCGGGAA	hhal/cfol nlaIII	acc651
		TTAATTGGGG GCAGGACCAT GGCTCTGAAT AACCTCTGAA AGAGGAACATT GTTCTAGGTAC CTTCTGAGGC CGAGAACCC ACCCTGAGAA	mn1I	mn1I
		GCTAGCCCT ATTAAGCCG CCTCTGGTAA CGGGACTTAA TTGGAGACTT	mn1I	ddeI acI
				acc651
				nsPBI
				bsaJI
2901	TGTTGTTGAG TTAGGGTGTG GAAGTCCCC AGGGTCCCC GCAGGCCAGA GTATGCCAGA CATTAGTCAG CATTAGTCAGTTC GTCAGTGTAA CTCAGTGTCTT	scrFI	ppu10I	bsaJI
		mvaI	nsII/availII	scrFI
		ecoriI	nlaIII	mvaI
		dsav	spHI	ecoRII
		bstNI	spHI	dsav
		apyI [dam+]	spHI	bstNI
		bsaJI	spHI	apyI [dcm+]
			spHI	bsaJI
3001	TGTTGTTGAG TTAGGGTGTG GAAGTCCCC AGGGTCCCC GCAGGCCAGA GTATGCCAGA CATTAGTCAG CATTAGTCAGTTC GTCAGTGTAA CTCAGTGTCTT	nlaIV	spfNI	bsaJI
		scrFI	ppu10I	bsaJI
		mvaI	nsII/availII	scrFI
		ecoriI	nlaIII	mvaI
		dsav	spHI	ecoriI
		bstNI	spHI	dsav
		apyI [dcm+]	spHI	bstNI
		ccgggtccc	rsPSPI	apyI [dcm+]
3101	CAGCAGGCCAG AAGTATGCAA AGCATGCCATC TCAATTAGTC AGCAACCATCA GTCCCCCCCC TAATCTCGCC CTAATCCGCC CTAATCCGCC GATTCGAGGG GTCTGTCGGTC	nlaIV	aciI	aciI
			aciI	aciI
			aciI	aciI
			aciI	aciI

FIG. 9K

3201 *bsrl acrl ccAGTTCGGC CCCATGGCT GACTAATT TTTTATTAT GCAGAGGGCC AGCCCGCTC GGCTCTGAG CTATTCCAGA AGTAGGTCT GATAAGGTCT TCATCACCG GGTCAAGGGC GGTAAAGGGC*

3301 *AGGGTTTTT GGAGGCCCTAG GCTTTTGCAA AAAGCTGTT CCTCGAGCGG CGCTTAATTC ATTAAATCC TGCAAGTAAAC AGCTTGGCAC TCCGAACAT CGAAACAGTT TTTCGACAAAT GGAGCAATTAA TCCCGGGGG TAAATTAGG AGTCCATTG TCGAACCTGG*

3401 *TGGCCGTGCGT TTACACAAGT CGTGACTGGG AAAACCCCTGG CGTTACCCAA CCTTAATCGGC TTGAGGACCA TCCCCCCTTC GCCAGCTGGC GTAATAGGA ACCGGCAGCA AAATGTTGCA GCACIGACCC TTTGGGACC GCAATGGGT GAATTAGGGG AACGCTGGT AGGGGGAG CATTATGCT*

nlariI
styI
ncoI
bsrl *acrl* **bsalI**
bsrl *ccAGTTCGGC* **CCCATGGCT** *GACTAATT* **TTTTATTAT** *GCAGAGGGCC* *AGCCCGCTC* *GGCTCTGAG* *CTATTCCAGA* *AGTAGGTCT* *GATAAGGTCT* *TCATCACCG* **GGTCAAGGGC** *GGTAAAGGGC*

3501 *AGAGGCCGGC ACCGATCGCC CTTCACAAACAA GTTGCGTAGC CTGAATGGCG ATAGGGCGCTT GATCCGGTAT TTTCCTCCTTA CGCATCTGT CGGTATTCA TCTCCGGGG TGGCTAGGG GAAAGGGTTGT CAACGCATCG GACTTACCGC AAAGAGGAAT GCTACCCATA AAAGGAGAC ACACCCGAC*

fnu4HI
afII *mnlI*
haelII/palI
bseJI *bgII* **ddeI**
haeIII/palI *bsaJI* *mnlI* *alul*
mnlI *mnlI* *acrl* *haelII/palI* *mnlI*
mnlI *mnlI* *aacl* *haelII/palI* *mnlI*

hinPI
hhAI/cfol

rmal
styI
bsaJI
blnl
avrII
haelII/palI
stul
haeI
mnlI *mael*
mnlI *mael*

fnu4HI
mcrI
eagI/xmaIII/ecXI
eaEI
bsrBI
xbaI *notI*
paer7I *haelII/palI*
avaI *fnu4HI* *paci*
maeIII *tagI* *cfrI*
mnlI *acrl* *acrl*
alul *mnlI* *acrl* *acrl*
mseI *ascI*

scfI
pstI
bsgI
bsaHII *ahaIII/draI*
swaI *sse8387I* *alul* *bsrI*

bspmI
bstU
bsh1236I
hinPI
hhAI/cfol *mseI*
bsaHII *ahaIII/draI* *maeIII*

scfI
pstI
bsgI
bsaHII *ahaIII/draI* *maeIII*

scfI
mvai
ecoriI
dsav
bstNI
apYI(dcm+)
bsaJI *maeIII*
mseI

fnu4HI
bbvI *fokI*

alul
pvuII
nsPvII
earI/ksp632I

mbolI
earI/ksp632I

hinPI
hhAI/cfol
nlarV
nari
kasi
hinII/acyI
hgICl
haelII *acrl*
banI *sfanI*
ahaII/bsaHII

sau3AI
mboI/ndeII [dam-]
dpnI [dam+]
dpnII [dam-]

sau96I
haelII/palI
asul
mnlI *acrl*

sfanI *acrl*

bgII
pvuII/bspCI
bsaHII

3501 *AGAGGCCGGC ACCGATCGCC CTTCACAAACAA GTTGCGTAGC CTGAATGGCG ATAGGGCGCTT GATCCGGTAT TTTCCTCCTTA CGCATCTGT CGGTATTCA TCTCCGGGG TGGCTAGGG GAAAGGGTTGT CAACGCATCG GACTTACCGC AAAGAGGAAT GCTACCCATA AAAGGAGAC ACACCCGAC*

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FIG. 9M

FIG. 9N

hg*i*AI/asPHI
bsp1286
bs*HK*AI
mb*l*/nd*l*[dam-]
dp*l*[dam+]
dp*l*[dam-]

sau3AI
mbo*l*/nd*l*[dam-]
dp*l*[dam+]
dp*l*[dam-]

ap*l*II/snoI
alw44I/snoI
mb*l*[dam-]
sf*AN*I
hphI

4601 CTGGTGAAG TAAAGATGC TGAAAGATCG TTGGGTGAC GAGTGGTTA CATCGAACTG GATCTAACAA GCCTGTAAGAT CCTTGAGAGT TTTCGCCCG GACCACCTTC ATTTCCTACG ACTTCATGC AACCCACGTG CTCACCAAT GTAGTTGAC CTAGAGTTGT CGGCCATTCTA GGAACTCTCA AAACCCGGC

nspBII
mbo*l*/nd*l*[dam-]
dp*l*[dam+]
dp*l*[dam-]

b*ST*YI/xho*l*
bs*r*I dp*l*[dam-]
al*W*I[dam-]
bst*Y*I/xho*l*
mb*o*II

scrFI
nc*l*
ms*p*I
hp*AI*
ds*A*V
ca*U*I
hin*l*/acy*l*
bst*U*I
bsh1236I
fnu*D*I/mvn*I*
hin*P*I
hh*AI*/c*f*o*I*
ah*AI*/bs*A*H
beg*I*
mc*R*I
f*nu*4H*I*
gc*GC*CATAACA
ac*l*I

ma*l*II
psp1406I
xmn*I*
asp700

4701 AAGAACGTTT TCCAAATGATG AGGACTTTA AAGTCTCTGT ATGGTGGCCG GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTG GCCCGGTTTCG GTGAGGCCAG CGCGGTATGT

ac*l*I
tha*I*
f*nu*4H*I*

rs*AI*
csp6I
sc*AI* h*phi*I ma*l*II
bs*AI*

4801 CTATTCTAG AATGACTTGG TTGAGTACTC ACCACTACA GAAAGCATC TTACGGATGG CATGACAGTA AGAGAMTTAT GGAGTGCTGC CATAACCAG GATAAGACTC TTACTGAAAC

sc*AI*
f*ok*I n*lai*II
f*ok*I n*lai*II
c*at*I

rs*AI*
csp6I
sc*AI* h*phi*I ma*l*II
bs*AI*

4901 AGTGATAACA CTGGGGCCA CTTACTCTG ACAACGATCG GAGGACCGAA GGAGCTAACCC GCTTMTTGC ACRACTGGG CCTCGATTGG CCTCTGGCTT CGAAAMACG TGTGTACAT TGAGGGAAAC

sa*u*96I
ava*l*
sau3AI as*u*I
mbo*l*/nd*l*[dam-]
dp*l*[dam+]
dp*l*[dam-]
p*v*u*l*/bs*p*CI
mc*RI* mn*l*I al*U* ac*l*I
fr*u*g*II*
ac*l*I

ma*l*III
n*lai*II
sau3AI
mbo*l*/nd*l*[dam-]
dp*l*[dam+]
dp*l*[dam-]
al*W*I[dam-]
p*v*u*l*/dam+
dp*l*[dam-]
actcgccrtg
ccttagtacat tgagggaaac

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FIG. 90

mspl
 hpaiI
 bswai
 nlarv alui
 rmai
 maeI
 5001 ATCGTGGGA ACGGAGCTG AATGAAGCCA TACCAAACGA CGAGGTGAC ACCACGATGC CAGCAGCAT CCCAACACG TTGCTGACTG TCGTGTAG TAGCAACCT TGGCTCGAC TTACTCGGT ATGGTTGCT GCTGCACTG AACGGTTG AACGGTGTG ATATTGACC

 mspl
 hpaiI
 scrFI
 alui ncII
 rmai dsav
 caulI
 maeI
 5101 CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTATAGAC TGGATGGAG CGGATAAAGT TGCAGGACCA CTTCCTGCCT CCGCCCTTCG GCCTGCCTGG TAATATCTG ACCTACCTTC GCCTTATTC ACGTCTGT GAAGACCGGA GCCGGGAGG CC3ACCGACC

 mspl
 hpaiI
 cfr10I
 nlaIV hphiI
 9sui/bpmI
 5201 TTATATGCTG ATAAATCTGG AGCCGGTAG CGTGGGTCTC GCGGTATCAT TGCAGGCACTG GGGCCAGATG GTAAAGCCCTC CCCTATGTA GTTATCTACA AAATAACGAC TATTTAGACC GCACCCACTC TGCGCCACTC GCACCCAGAG CGCCATAGTA ACGTCTGTAG GCCTATGCTGCTAC CTTCTGGAG CCCTGGGTAC CTTCTGGAG CCCTGGGTAC CTTCTGGAG CCCTGGGTAC

 mspl
 hpaiI
 fokI
 mseI
 asel/asnl/vspI
 mnlI
 5301 CGACCCCTC AGTCCGTTGA TACCTACTTG CTTATCTG CTAGCGACTC TATCCACGGA GTGACTAATT CGTAAACATT CGTAAACGGA GACAGCTGG TTCAATGAG

 mspl
 hpaiI
 fokI
 mseI
 5401 ATATATACCT TAGATGATC TAAACTICA TTCTTAATT AAAAGGACT AGGTGAAGAT CCTTTTGTGAAAT CTTCTGATGA CCAAAATCCC TTAAACGTGAG TATATATGAA ATCTTAACCTAA ATTTTGAGT AAAAATTTAA TTCTCTCTAGA TCCACTCTCA GGAAAAGACTA TTAGAGTACT GTTTTGTAGGG AAATTCGACT

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FIG. 9

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FIG. 9G

FIG. 9R

thaI
 fnuDII/mvnI
 bstUI
 bsh1236I
 hinPI
 hhaI/cfoI
 thaI
 fnuDII/mvnI
 bstUI
 bsh1236I haell/palI
 bshI eaeI tfII aseI/asnI/vspI
 nspBII
 bsrI
 acII
 cfrI
 hinfl mseI
 nspBII
 6301 AGTGAGCGAG AAAGGGAAAT AGCGCCCAAT ACGCCAAACCG CCTCTCCCCG CGCGTGGCC GATTICATTA TCCAGCTGGC ACGACAGGTT TCCCGACTGG
 TCACTCGCCTC CTTCGCCTC TCGGGTAA TGCGGTAA TGCGTGGTCCAA AGGGCTGACC
 scrFI
 mvaI
 ecoRII
 dsAV
 nlaIV bstNI
 hgIC1 apyI (dcm⁺)
 msplI
 hpall
 bani
 bsAJI
 GCTTTACACT TTATGCTTCC GCCTCGTANG TTGTGTTGAA
 GGCACCCAG CCGGGGTC CGAATGTGA AATACTGAGG CCGAGCATAAC AACACACCTT
 tru9I
 maeI
 aseI/asnI/vspI
 xmnI
 acII
 bstRI
 6401 AAAGGGGCA GTGAGCCAA CGCAATTAAAT GAGGTTAAC TCACITCARTA GGCACCCAG GCTTTACACT TTATGCTTCC GCCTCGTANG TTGTGTTGAA
 TTTGGCCGT CACTCGCTT GCGTTAAATT CACTCAATGG AGTGAGTAAAT CGGTTAAAT CACTCAATGG AACACACCTT
 tru9I
 maeI
 aseI/asnI/vspI
 xmnI
 acII
 bstRI
 6501 TTGTGAGCGG ATAACACATT CAACAGGAA ACAGCTATGA CCATGATAC GAATTAA
 AACACTCGCC TATTTGTTAA GTGTGCTT GTGTGATACT GGTTACTAATG CTTAATT

> length: 6557

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FIG. 10A

1 TTCTGAGCTCG CCCGACATTC ATTATTGACT AGTTTAAAT AGTAATCAT TACGGGGTCA TTAGTCATA GCCCATATAT GGAGTTCGC GCATACATAC
 AAGCTGAGC GGGCTGAAAC TAATACGTGA TCATAATTA TCATTAGTTA ATGCCCCAGT ATCAAGTAT CGGGTATATA CCTCAGGGG CAATCTATTG

scrFI
 mvaI
 ecorII
 dsav
 acII
 bglI bstNI
 sau96I
 haeIII/palI acII
 asuI apyI [dcm⁺] acI
 101 TTACGGTAA TGCCCGGCCGCT GGCTGACCGC CCAACGACCC CGGCCCATGG ACGTCAATAA TGACTATGT TCCCATAGTA ACGCCATAG CCTGATTCGA
 AATGCCATT ACCGGGGGA CGGACTGGCG GGTTCCTGGGG GGGGGGTAAAC TGCACTTATT ACTGATACA AGGGTATCAT TGCCTTATC CCTGAAAGGT

maeII
 hinII/acyI
 ahaII/bsaHII
 aatII
 bgI I
 rsal
 csp6I
 ndel
 201 TTGACGTCAA TGGGGAGT ATTACGGTA AACTGCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA ATGACCCCC CTATTGACGT CAAATGACGT
 AACTGCCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG ACCGGTCA TAGTTCACAT AGTATACGGT TCATGCCGG GATACTGCA GTTACTGCCA

scrFI
 mvaI
 ecorII
 acII
 bglI dsav
 sau96I bstNI
 haeIII/palI
 asuI apyI [dcm⁺] acI
 301 AAATGGCCG CCTGGCAATA TGCCCAGTAC ATGACCCAT GGGACCTAT TACCTGGAG TACATCTACG TATTAGTCAT CGCTTATTACG ATTAGATGC
 TTACCGGGC GGACCGTAAT ACGGGTCA TGACTGAAATA CCCTGAAGG ATGAACCGTC ATGAGATGC ATAATCAGTA GCGATAATGG TACCACTAG

thaI
 fnuDII/mvnJ
 bstUI
 bsh1236I
 acII maeII
 bsII
 rmaI
 mseI
 speI aseI/asnI/vspI
 rmaI
 maeI
 tru9I
 maeII
 acII
 maeII
 hinII/acyI
 ahaII/bsaHII
 aatII
 maeII
 maeII
 hinII/acyI
 ahaII/bsaHII
 aatII
 maeII
 rsal
 csp6I
 ndel
 maeII
 hinII/acyI
 ahaII/bsaHII
 aatII
 maeII
 rsal
 csp6I
 ndel
 maeII
 nciI
 dsal bphI acII
 bseJI sfaNI
 styI
 nlaIII
 maeII
 snal
 bsaAI
 csp6I
 maeII
 rsal
 csp6I
 nlaIII
 styI
 nlaIII

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FIG. 10B

19. TUB
 401 GGTGGCA GTACATCAT GGGGGGAT AGCGGTTGA CTCACGGGA TTCCAAAGTC TAGGCCCTT AAAGGTTCAAG AGCTGGGTTA ACTGGAGTTA CCCCTCAAACAA AACCCCTGTT
 CAAAAACCGT CATGTAGTT CCCGCACCTA TCGCCAAACT CGTGCACCTA hinlli/acyl
 rsal aluI
 csp6I sstI
 acII sacI
 hinfl hgjII
 hgiII aspH
 acII ec1136II
 bsp1286 bsrKAI
 bmyI banII
 rsal haeIII/pali
 acII csp6I mnlI
 hgai AAATGGGGC TAGGGGTGA CGGTRGGAGG TCTATATAAG CAGATATTG GTCRCGAGCA
 maeIII acII csp6I mnlI
 AAATGGGGC TAGGGGTGA CGGTRGGAGG TCTATATAAG CAGATATTG GTCRCGAGCA
 501 AAATCAACGG GACTTCCAA AATGTCGTA CAACTCGCC CCATTGACGC AAATGGGGC TAGGGGTGA CGGTRGGAGG TCTATATAAG CAGATATTG GTCRCGAGCA
 TTAGTTGCC CTGAAAGGTT TTACAGCATT GTTGGGGGG GTTAACCTGGG
 esp3I
 scrfI
 mvaI bsmAI
 ecorII dsaV
 bstNI bsrKAI
 apyI [dcm+]
 sau3AI gsuI/bpmI
 mboI/ndeI [dam-] hgaI fokI
 dpmI [dam+] dsaV
 dpmII [dam-] ahaiI/bsaHI mnII
 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTGGACCT CCATAGAAA CACCGGACG GATCCAGGCCT CGGCCCTGG CTAAGTCTCT
 BACTACTGG CAGCTTAGGG GACCTCTGG STAGSTGGA CAAAACTGGA
 601

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FIG. 10C

tfiI acII
 thal hinfi fnuDII/mvnl bstU bsh1236I
 acII rsAI scfI csp6I scfI hinfi asuI
 maelli rscfI pleI scfI haelli/pali
 bstXI sau96I styI
 bstU/mvnI tru91
 bsh1236I asuI/asnI/vspI
 701 TTAGAACCGG GATTCCCCGT GCCAAGACTG ACGTAAAGTAC CGCTATAGCA GCTATAGGC CCACCCCTT GGCTTCGTTA GAACGGGCT ACAATAATA
 AACCTGGCG CTAAAGGCGA CGGTTCAC TGCATTCATG GCGATATCT CAGATATCCG GGTGGCGAA CGGAAGCAA TTTCGGCGA TTGTTAATTAT
 sau96I avall asuI
 scrPI mvnI
 ecORII dsav
 bstNI apyI [dcm+]
 bsuII bsuJI
 maeIII fokI
 hphI scfI
 801 CATAACCTTA TGATCATAC ACATAGCATT TAGGTGACAC TATAGATAA CATCCACTTT GCCTTTCTT CCACAGGTGT CCACCTCCAG GTCCCAACTGC
 GTATTGGAAAT ACATAGTATG TGTATGCTAA ATCCACTGTG ATTATCTATT GTAGGTGAAA CGGAAAGAGA GGTTTCACCA CAGGTGACG
 hinII/acyI ahall/bsaHI
 aatII
 thaI
 fnuDII/mvnI bstU
 acII maeII hphi bsh1236I tagI
 aluI taqI hindIII
 mnII
 bsaJI ddeI clai/bsp106
 901 ACCTCGGTTC TAAGCTTATC GATATGAAAA AGCCTGAACT CACCCGACG TCGTGTGAGA AGTTTCGAGA CGAAAGCTGAT CGAACGGCTCT CGGACCCGTAT
 TGGAGCCAG ATTCAATAAG CTATACTTT TCGGACTTGA GTGGACTTGA AGACAGCTT TCAAAGACTA GCTTTCAGA CTGTGGAGA GGCTGGACTA
 hinPI
 aluI tfiI
 fnu4HI hinfi
 mnII aluI
 bbVI mnII mbolI aluI aluI hhaI/cfoI
 1001 GCAGCTCTG GAGGGAAAG AATCTGTGC TTTCAAGTTC GATGTTAGGAG GGGGTGGATA TGTCCGTGGG GAAATAGGT CGCCCGATGG TTTCATACAA
 CGTCGAGAGC CTCCGCTTC TTAGGACAG AAAGTCGAG CTACATCCTC CGGCACCTAT ACAGGACGCC CATTATCGA CGCGCTTACCG AAAGATGTTT

FIG. 10D

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hinPI	hhAI/cfol	msPI	
	thaI	hpAI	
	mroI		
acII	haeIII/palI	bspMII	
	mcRI fnuDII/mvNI	bsPEI	
sau3AI	bsPEI/xmaIII/ecII	bsaWI	
mboI/ndeII [dam-]	eaEI bstUI	tfII	
dpnI [dam+]	cfrI bsh1236I	hinfl	
dpnII [dam-]	sfaNI fnu4HI	bsII acIII	
1101 GATCGTATC TTATCGCA CGCCGCTCC CGATTCGGGA AGTGCCTGAC ATTGGGAAT TCAGGGAGAG CCTGACCTAT TGCAGCTCC		apoI	sfANI acII
	GCTAAGGCC		
	GGGGTAGAC		
	AAATAGCCGT		
	CTAGCAATAC		
hgiAI/asPHI	styI		
bsp1286	thaI ncoI		
bsIKKAI	fnuDII/mvNI		
bmyI	bbVI mcrI	haeIII/palI	
apaLI/snoI	mspI bsh1236I	bsaJI sfaNI	
alw44I/snoI	pstI hpaII	mnII nlaIII	
bsII draIII	acII bsgI	frl0I acII haeI	
1201 GCGGTGACA GGGTGTACG TTGCAACACC TGCTGAAAC CGAAACTGCC GCTGTTCTGC AGCCGGCATG GATGGATCG CTGGGCCGA	bspMI	fokI mcrI bbVI cfrI dpnII [dam-]	
		ctacgctac	
		ctccggta	
		gacggccgt	
sau96I	thaI		
avaII	fnuDII/mvNI	nlaIII	
asul	bstUI	mboI/ndeII [dam-]	
	bsph1236I	dpnII [dam+]	
acII	hinPI	dpnII [dam-]	
bsrBI	bsaJI	alwI [dam-]	
ddeI	haeIII/palI		
1301 TCTTAGCCAG ACGAGCCGGT TCGGCCATT CGAACCCCA GGAATCGGTIC AATAACACTAC ATGGGTGAT TTATATGG CGATTGGCTGA TCCCAGTG	tffI		
	hinPI		
	hhaI/cfol		
	thaI		
	fnuDII/mvNI		
	bstUI		
	bsaJI		
	haeIII/palI		
	sau96I		
	asul mnII		
	bsII		
	draIII		
tth111I/asPI	hgaI		
drdI	ccgtcgccgt		
bsrI	ccgtcgccgt		
1401 TATCACTGGC AACCTGTGAT GGACGACACC GTGAGTGGT GAGCTCTGAT GAGCTGATGC TTGGGGCCGA GGACTTCCCC GAAGTCGGC	tttgcgtgg		
	AGAATGGTC TGCTGGCCA AGCCGGTA		
	CCCTGGCTT CCTAGCCAG TTATGTGATG TACCGACTA AAGTATAAGC GCTAACGACT AGGGTACAC		

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FIG. 10E

1501 acii
 thaI
 fnuDII/mvnI
 hgIAI/asPHI
 bsp1286
 bsiIKAI
 bmyI bstUI
 apalI/snoI
 alw44I/snoI
 mnII
 bsh1236I nlaIV
 CGGGATTTGCA CGCTCCAACA ATGTCCTGAC GGACATAGGC CGATAAACAG CGGTCATGA CTGGAGCGAG GCGATGTTGG GGGATTCCCA
 TGGAGCACGT GGCCTAAAG CCGAGGTGT TACAGGACTG CCTGTTACCG GGTATTGTC GCCAGTAACT GACCTCGTGC CGCTACAAGC CCCTAAAGGT
 fnu4HI

1601 acii
 fnu4HI
 haelli/pali
 eaeI
 cfri
 nspBII
 acii
 nspBII
 sacII/stII
 nspBII
 kspI
 dsal
 bsajI
 acii
 fnu4HI
 fnuDII/mvnI
 bstUI
 bsh1236I
 sacII/stII
 nspBII
 msPI
 hpall
 mroI
 bspMI
 bspEI
 bsawI
 sau3AI acii
 mboI/ndeII[dam-]
 dpnII[dam+]
 dpnII[dam-]
 alwI[dam-]
 dsal
 haelli/pali
 mboII mnII bsajI
 mboII gsul/bpmI
 mnII
 ATACGAGGTC GCCAACACCT TCTTCCTGGAG GCGCTGGTGG AGCTTGATGG ACCAGCAGAC GTACTTCAG CGGAGGCATC CGAGGCTTC AGGATGCCG
 TCTGCTCCAG CGGTCTGAG AGAAGACCTC CGGGACCAAC CGAACATACC TCTGCTGCTG CAGAACCTC GCCTCCGTAG GCCTCGAAG TCCTACGGG
 scrFI
 ncII
 msPI
 hpall
 dsav
 cauII
 nlaIV
 1701 CGGCTCCGG CGTATATGCT CGGCATTGGT CTTGACCAAC TCTATCAGAG CCCAATTTCG ATGATGCAAG TTGGTGGTGCAG GGTCTGTCAG GGTCTGTCG
 GCCGAGGCC GCATATACGA GGGTAAACCA GAACCAACTG CCGTTAAAGC TACTACGTC AGATGTTG AGATAGTCTC GAACCAACTG AACCCGCGTC CCAGCTACGC

alwI
 fnu4HI
 bbVI
 hinPI
 hhal/cfol sfaNI
 alwI
 nincII/hindII taqI sfaNI
 hinPI
 taqI
 drdI
 hgAI

FIG. 10F

1801 ACGCAATCTGT CCGATCCGGA GCGGGGACTG TCGGGGGTAC ACAAATGCC CGCAGAAGGC GGGCGTCTG GACCGATGGC TGTGTTAGAAG TACTGCCGA
 TGCGTTAGCA GGCTAGGCT CGGCCCTGAC AGCCCCGATG

nlaIV msPI hpII bsII mroI bspMII bspEI [dam-] bsAWI accIII [dam-] sau3AI mboI / ndeII [dam-] dpnII [dam+] dpnII [dam-] alwI [dam-]	scrFI ncII msPI hpII dsAV xmaI / pspAI scrFI ncII dsAV cauII bsAJI avAI bsAJI	haeIII / pali mcRI eagi / xmaIII / ec1XI eaeI cfrI fnu4HI acII chaI fnuDII / mvnI bstUI bsh1236I hinPI hhaI / cfoI asuI scAI rsal csp6I sau96I avail asuI scAI scrFI ncII msPI hpII dsAV xmaI / pspAI scrFI ncII dsAV cauII bsAJI avAI bsAJI	sau3AI mboI / ndeII [dam-] dpnII [dam+] dpnII [dam-] alwI [dam-]	nlaIV cauII bstVI / xholI bamHI bsAJI alwI [dam-]	acII fnu4HI asuI bgII nlalII sfII styI easI ncoI cfrI dsAI tagI haeIII / pali clal / bsp106 bsAJI	sau96I haelli / pali bsII sfANI hhaI / acyI hgaI alwII / bsaHI	1901 TAGTGAAAC CGACGCCCA GCACTCGTC GAGGGCAAG GAATAGAGTA GATGCCGACC GAAGGTCCC CGGGGAATTG AATCGATGCC ATCCATCTTACGG CTACGGCTGG CTCCTAGGG CCGCTACCG ATCACCTTG GCTGGGGGT CGTGAGCAGG
---	---	---	--	--	---	--	--

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FIG. 10G

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FIG. 10H

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FIG. 101

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FIG. 10J

	mspI hpAI bsII bsAII	sau3AI mboI/ndeII [dam-] dpnI [dam+]
scrFI mvaI ecORII dsav bstN1 apY1 [dcm+]	gsuI/bpmI scrFI mvaI haeIII/pallI fnu4HI thaI mnII fnuDII/mvNI bstU1 bsh1236I 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC	ecorII mvaI haeIII/pallI fnu4HI thaI mnII bstNI apyI [dcm+] bstU1 bsh1236I 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
	acII bstNI apY1 [dcm+]	acII bstNI apY1 [dcm+] pleI rsal csp6I hinfi 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
		pleI rsal csp6I hinfi 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
		gstI mboI/ndeII [dam-] dpnI [dam+] dpnI [dam-] alwI [dam-] nlaIV bstYI/xholI bamHI alwI [dam-] 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
		gstI mboI/ndeII [dam-] dpnI [dam+] dpnI [dam-] alwI [dam-] nlaIV bstYI/xholI bamHI rsal mnII alwI [dam-] acc65I 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
		gstI mboI/ndeII [dam-] dpnI [dam+] dpnI [dam-] alwI [dam-] nlaIV bstYI/xholI bamHI rsal mnII alwI [dam-] acc65I 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
		gstI mboI/ndeII [dam-] dpnI [dam+] dpnI [dam-] alwI [dam-] nlaIV bstYI/xholI bamHI rsal mnII alwI [dam-] acc65I 3001 TATCAACAGA AACCAGAAA AGCTCCGAA CTACTGATT AGCGGGCCTC
		scrFI mvaI ecorII dsav bstN1 apY1 [dcm+]
		scrFI mvaI haeIII/pallI haeI mnII asp700 3301 CTGAATAACT TCTATCCCAG AGAGGCCAA GTACAGTGG AGTATTGCTC
		scrFI mvaI haeIII/pallI haeI mnII csp6I 3301 CTGAATAACT TCTATCCCAG AGAGGCCAA GTACAGTGG AGTATTGCTC
		scrFI mvaI haeIII/pallI haeI mnII csp6I 3301 CTGAATAACT TCTATCCCAG AGAGGCCAA GTACAGTGG AGTATTGCTC

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FIG. 10K

sstI
 sacI
 hgjII
 hgIAI/asPHI
 ec1136II
 bsp1286
 bsiWKAII
 bmyI
 haelli/pali
 sau96I aluI
 asuI banII
 ddeI eco0109I/draII
 celIII/espI
 bpu1102I
 hgaI
 accI
 hphI
 maeIII
 alwNI ddeI
 3401 AGGACAGCAC SCFI mnII bbVI TGACAGGCC CAAAGCAGAC TAGGAGAAC ACAAAAGTCTA CGCCCTGGAA GTCACCCATC AGGGCTGAG TCGTGTGGG ACTGGGACTC GTGTCGGAG TCGTGTGGTG ATGTCGTCTG ATGCTCTTTG CAGTGGTAG TCCGGACTC
 sau96I
 nlalII
 acI haelli/pali
 fnu4HI asuI
 bgII styI
 aluI sfII ncoI
 hindIII eaeI dsalI
 truGI cfrI bsajI
 maeIII tagI haelli/pali
 mseI bbVI
 maeIII aluI CGTTCGATGG CGGCCATGGC CCAACTGTT TATTGCAGCT TATAATGGTT ACAAAATAAAG
 3501 CTCGCCGTC ACAAAGAGCT TCAACAGGG AGAGGTAA GAGGTCTCC TCTCACAAATT CGAAGCTAC GGGGTACCG GGTGAAACAA ATAACGTCGA ATATTACCA ATTTACCA TGTTTATTC
 sauJAI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 pvuI/bspCI
 mcrI
 taqI(dam-)
 clai/bsp106(dam-)
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 rmaI
 bsmI maeI
 sfaNI apoI
 3601 CAATAGCATC ACAAATTCA CAATAAAGC ATTTCATCA CTGCATTCTA GTTGTGGTT GTCCAACACT ATCAATGAT CTTATCATGT CTGGATCGAT GTTATCGTAG TTGTTAAAGT GACGTAAGAT TAAAAAAGT GAGTTACATA GAATAGTACA GACCTAGCTA

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FIG. 10L

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FIG. 10M

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FIG. 10N

FIG. 10N
 mnII
 nlaIV
 hgiJI
 bsp1286
 bmyI
 banII
 nlaIV
 hgiCI
 banI
 taqI
 hphiI
 cfr101
 aluI
 GCCGGTTC CCCGTCAGC TCTTAATTCGG GGGCTCCCT TAGGGTCCG ATTASTGCT TTACGCC ACCGACCCAA AAAACTGAT TTGGGTGATG
 4501 CGTCACGTAG TGGGCCATCG CCGTGTAGA CGGTTTTTG CCCTTGACG TTGGAGTCGA CGTCTTAA TAGTGGACTC TTGTTCAAA CTGGAACAC
 CAAGTGCATC ACCCGTAGC GGGACTATCT GCCRAAAAGC GGAAAACTGC AACCTCAGGT GCAAGAAATT ATCACCTGAG AACAAGTTT GACCTGGTGC
 maeII
 haeIII/palI
 draIII
 sau96I
 bsaAI
 asuI
 4601 ACTCAACCCT ATCTGGGT ATTCTTGA TTATAAGGG ATTGGCCGA TTTCGGCTA TTGGTTAAA ATAGAGCTGA TTAAACAAAA ATTAAACGGC
 TGAGTGGGA TAGAGCCCCGA TAAGAAACT AAATATTCCC AAACCCGAT AACCAATT TTACTCGACT AAATTGTTT TAATTGGCGC
 mspl
 hpaII
 nael
 cfr101
 GCTGGTATG GCTGGCCCC GACACCCGGC AACACCCGGT GACGGGGCTT GCTGACCGTC GCTCCCCAAC CTCGGGGGA TTGTGGGG
 4701 AGATTAAC AAATTAAC GTTACATT TTATGTCGA CTCTCAGTAC AATCTGCTT GATGCCCAT AGTTAAGCCA ACTCCCTAT CGCTACCTGA
 TTAAATTGCT TTATAATTG CAATGTTAA AATACACGT GAGAGTCATG TTAGACGAGA CTACGGCGTA TCAATTGGGT TGAGGGATA GCGATCCACT
 tru9I
 bsII
 avaiI
 4701 ACTCAACCCT ATCTGGGT ATTCTTGA TTATAAGGG ATTGGCCGA TTTCGGCTA TTGGTTAAA ATAGAGCTGA TTAAACAAAA ATTAAACGGC
 TGAGTGGGA TAGAGCCCCGA TAAGAAACT AAATATTCCC AAACCCGAT AACCAATT TTACTCGACT AAATTGTTT TAATTGGCGC
 hg1AI/asPHI
 bsp1286
 bshKAI
 maeII
 bmyI
 ddeI
 apaLI/snoI rsI
 alw44I/snoI csp6I
 4801 AAATTAAC AAATTAAC GTTACATT TTATGTCGA CTCTCAGTAC AATCTGCTT GATGCCCAT AGTTAAGCCA ACTCCCTAT CGCTACCTGA
 TTAAATTGCT TTATAATTG CAATGTTAA AATACACGT GAGAGTCATG TTAGACGAGA CTACGGCGTA TCAATTGGGT TGAGGGATA GCGATCCACT
 hinPI
 hhaI/cfoI
 thaI
 fnuDI/mnVI
 bstUI
 nspBII
 bsh1236I
 acII
 hgaI
 4901 CTGGTATG GCTGGCCCC GACACCCGGC AACACCCGGT GACGGGGCTT GCTGACCGTC GCTCCCCAAC CTCGGGGGA TTGTGGGG
 GACCCAGTAC CGACGGGGGG CGACGGTAC CGACGGGGGG

FIG. 100

scrFI thaI
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI

nspI nspI
hpAI fnu4HI
dsAV aluI nspHI
cauI bbvI nlaiI
5001 TCCGGGAGCT GCATGTTCA GAGGTTCAC CGGTATCAC CGAACCGGC GAGGAGTAT TCTTGAAAC GAAAGGGCCT CGTGATAACG CTATTTAT AGGCCCTCGA CGTACACAGT CTCCAAAAGT GGCAGTAGTG GCTTGTGGC CTCGGTCATA AGAACCTCTG CTTTCCCGA GCACATGGC GATAAAATA

nlari acI
thaI
fnuDII/mvnI
bstUI
bsh1236I

maelI hincII/acYI
hinPI
bsh1236I

tru9I rcaI hincII/bsaHI
ddeI atIII
mseI bspHI
5101 AGGTTAATGT CATGATAATA ATGGTTCTT AGACGTCAGG TGGCACTTT CGGGAAATG TGCGGGAAAC CCCTATTGT TTATTTCT AAATACATC TCCAAATTACA GTACTATTAT TACCAAGAA TCTGCAGTC ACCGTAAAAA GCCCCTTAC ACGGCCTTG GGGATAAACAA AATAAAAGA TTATGTAAG

rcaI
bspHI
bsrBI bsmAI
aci nlaiI
5201 AAATATGTT CGGCTCATGA GACAATAACC CTGATAATG CTTCAATAAT ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTCCGTGT CGCCCTTATT TTTTACATA GGGGAGTACT CTGTATTGG GACTATTAC TAACTTTCT CTTCTCATAC TCATAAGTTG TAAGGCACA GGGGAAATAA

hgIAI/asphi
bsp1286
sau3AI
bsiHKA
mboI/ndeII(dam-)
dpnI(dam+) bmyI
dpnI(dam-)
eco57I
apaLI/snoI

fnu4HI
aciI
5301 CCCCTTTTG CGGCATTG CCTTCCTGTT TTGCTCACC CAGAAACGCT hphiI hphiI
GGTGAAGTA AAAGATGCTG AAGATCAGTT GGTCGACGA GTGGGTTACA GGGAAAAAC GCCGTAAC GGAAGGACAA AACAGTGGG GCTTGTGGC CCACCTCAT TTCTAGTCAA CCCACGTGT CACCCAAATG

FIG. 10P

sau3AI
 mboI/ndelI [dam-] sau3AI
 dpnII [dam+] mboI/ndelI [dam-]
 dpnII [dam-] dpnII [dam+]
 bstYI/xholI dpnII [dam-]
 bsrl nsPBII alwI [dam-]
 tagI alwI [dam-] acII bstYI/xholI
 5401 TCGAACTTGA TCTCAACAGC GGTAAATGCC TTAGAGTTT CGGCCCCCAA GAACGTTTC CAATGATGAG CACTTAAAG CTTGGCTT AACTCTCAA AGCTGACCT AGAGTTGTCG CCATTCTAGG AACTCTCAA
 scrFI
 nciI
 mspl
 hpaII
 dsav
 cauI
 hinII/acyI
 hgaI
 ahdII/bsaHI bcgI mcrl fnu4HI
 5501 ATTATCCCGT GATGACGCG GGCAAGAGCA ACTCGGTGCG CGCATACACT ATTCTAGAA TOACTTGGGT GAGTACTCAC CAGTCACAGA MAAGCATCT TAATAGGGCA CTACTGGGC CGGTTCCTCGT TGAGGCCAGG GCGTATGTA TAAGAGTCTT ACTGAGTCT ACTGAACCA CTCATGAGTG GTCAAGTGTCT TTTCGTGAA
 acII
 ddeI
 mcrl fnu4HI
 5601 ACGGATGGCA TGCAGCTTGA AGAATTATGC AGTGTGTCGA TAACCATGAG TGATAACACT GCGGCCCAACT TACTTGTGAC AACGATGGA CGCCGGTGA ATGAGAGCTG ACTATGTC ACTGGTACTC ATTGGTACTC
 nlaiI
 fnu4HI bbvI nlaiI
 TGCTTACCGT ACTGTCAATC TCCTTAATCG TCACGACGGT ATTGGTACTC ACTGGTACTC
 nlaiI
 sau3AI maelliI
 mboI/ndelI [dam-] sau3AI nlaiI
 dpnI [dam+] mboI/ndelI [dam-]
 alwI [dam-] dpnI [dam+]
 nlaiI dpnII [dam-]
 aluI acII nlaiI dpnII [dam-]
 5701 AGCTAACCGC TTTTTGGAC AACATGGGG ARCATGTAAC TCGCCTTGAT CGTGGAAC CGGAGCTGAA TGAAGCCATA CCAACGAGC AGCGTGAAC
 TCGATTGGCC AAAAACGTC TTGTACCCCC TAGTACATTC AGCGGAACTA GCACCCCTTG CCTTGGACTT ACTTGGTAT GTTGTGTC AGCGACTG

FIG. 10Q

5801 CACGATGCCA GCAGGAATGG CAACAACTT GCGCAAACCA TTAACCTAC AGTACCTTGTTGTTGAA CGCGTGTACCGT GTGCTACGGT ATTATCTGAC CTACCTCCGC
 mspl mspI
 hpaII hpaII
 scrFI scrFI
 aluI ncII tru9I fokI
 avilI fspI bsrlI mseI bsrlI acII
 maeII hhaiI/cfoI tru9I dsaV maeI caulI aseI/asnlI/vspI mnII
 PSP1406I
 bbvI bbvI
 sfaNI sfaNI
 5901 GATAAAGTTG CAGGACCCT TCTGCCTCG GCCTTCGCG CTGGCTGGTT TATTGCTGAT AAATCTGGAG CGGGTGAAGC TGGCTCTGC GSTATCATTC
 mspl mspI
 hpaII hpaII
 cfr10I
 nlalV hphI gsul/bpmI
 mspl
 hpaII
 hhaI/cfoI
 sau96I haelII/palI mspl
 avalI haelII/palI mspl
 asul haelII/palI mspl
 6001 CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACAG ACGGGAGTC AGGCAACTAT GGATGAAGCA ATAGACAGA TGGCTGAGT
 GTCCGTGACCC CGGTCTACCA TTGGGGAGGG CATAGCTCA CCTACTGCT CCTACTGCT AGGCACTTC TTATCTGCT ACCAGAGGC CCATAGTAAAC
 mspl mspI
 hpaII hpaII
 scrFI scrFI
 aluI ncII tru9I fokI
 mspl mspI
 hpaII hpaII
 cfr10I
 nlalV hphI gsul/bpmI
 mspl
 hpaII
 hhaI/cfoI
 sau96I haelII/palI mspl
 avalI haelII/palI mspl
 asul haelII/palI mspl
 mnII
 nlalV
 bsrlI haelII/palI mnII
 6101 AGGTGCCTCA CTGATTAAGC ATGGTAAC GTCAAGACCA GTTACTCAT ATATACCTTA GATTGATTAA AACTCTATT TTTAATTAA AGGATCTAG
 TCCACGGAGT GACTAATTG TAACCATTGA CAGTCTGGT CAATGAGTA TATAGAAT CTAACAAAT TTGGAAGTA AAATTAATT TTCTAGATC
 mspl mspI
 hpaII hpaII
 scrFI scrFI
 aluI ncII tru9I fokI
 mspl mspI
 hpaII hpaII
 cfr10I
 nlalV hphI gsul/bpmI
 mspl
 hpaII
 hhaI/cfoI
 sau96I haelII/palI mspl
 avalI haelII/palI mspl
 asul haelII/palI mspl
 mnII
 nlalV
 hgicI hgicI
 banI banI
 6101 AGGTGCCTCA CTGATTAAGC ATGGTAAC GTCAAGACCA GTTACTCAT ATATACCTTA GATTGATTAA AACTCTATT TTTAATTAA AGGATCTAG
 TCCACGGAGT GACTAATTG TAACCATTGA CAGTCTGGT CAATGAGTA TATAGAAT CTAACAAAT TTGGAAGTA AAATTAATT TTCTAGATC

FIG. 10R

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FIG. 10S

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FIG. 10T

FIG. 10T
 7001 GTGAGCTGAT ACCGGTCGCC GCAGGCCAAC GACCGAGGC AGCCGAGTCAG TGAGCGAGGA AGCGGAAGAG CGCCCAATAAC GCAAACGCC AGCGCCCGG
 CACTCGACTA TGGCGAGGG CGTGGGCTTG CTCGCTCGG CTCGCTCTC ACTCGCTCAGTC TCGCTCTCTC ACTCGCTCTC AGCTGGTTATG CGGCGTTTC GCGCTCTCTC AGAGGGGGC
 bsrBI acII fnu4HI bbVI
 aluI acII fnu4HI mcrI
 7101 CGTTGGCGGA TTCAATTAAATC CAGCTGGCAC GACAGGTTC CCGACTGGAA AGCGGGCACT GAGGCCAACG CATTAAATGT GAGTAACTTC ACTCATTAAGG
 GCAACGGGCT AAGTAATTAG GTCGACCGGTG CTGTCCAAG GGCTGACCTT TCCTCCCTCA CTGCGCTTC TGTAATTACA CTCATGGAG TGACTAATCC
 scrFI mvaI ecoRII dsBAV bstNI
 apyI [dcm+] msP1
 bsAI I hpaII
 7201 CACCCCCGGC TTACACTT ATGCTTCCGG CTCTGTGTT GTCGCTGATT GTCGCTGATT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA
 CTGGGGTCCG AAATGTAAA TAGATGAA GAGGATACAA CACACCTAA

>length: 7305

INTERNATIONAL SEARCH REPORT

Inte: onal Application No
PC1/US 95/09576

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;, pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors' see page 192, line 7 - page 196, line 5; figures 2,3	1-3,7,8
Y	---	5,6, 9-12, 16-21
	-/-	

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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- *'E' earlier document but published on or after the international filing date
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- *'P' document published prior to the international filing date but later than the priority date claimed

- *'T' later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- *'&' document member of the same patent family

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Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Fax (+31-70) 340-3016

Authorized officer

Hornig, H

INTERNATIONAL SEARCH REPORT

Int'l	Serial Application No
PCT/US 95/09576	

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CELL, vol. 37, no. 3, July 1984 CELL PRESS, CAMBRIDGE, MA, US;, pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application pZIP-Neo SV(B)1 see figure 1 ---	1-3, 7, 8
Y	MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC, US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1 ---	5, 6, 9-12, 16-21
Y	WO,A,94 05784 (US) 17 March 1994 see the whole document ---	5, 6, 9-12, 16-21
Y	EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987 see the whole document ---	5, 6, 9-12, 16-21
A	WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document ---	1-21
A	WO,A,90 12025 (UNIV LELAND STANFORD JUNIOR) 18 October 1990 cited in the application see the whole document ---	1-21
A	EP,A,0 260 148 (GENENTECH INC) 16 March 1988 see the whole document ---	1-21
A	EP,A,0 160 457 (GENENTECH INC) 6 November 1985 cited in the application see the whole document ----	1-21
2		-/-

INTERNATIONAL SEARCH REPORT

Int'l Application No
PCT/US 95/09576

C(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL.ACAD SCI., vol. 86, February 1989 NATL. ACAD SCI., WASHINGTON, DC, US;, pages 1041-1045, M. VIVAUD ET AL. 'A 5' splice-region G-C mutation in exon 1 of the human beta-globin gene inhibits pre-mRNA splicing: A mechanism for beta-thalassemia' see the whole document -----</p>	1-4

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
PCT/US 95/09576

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9405784	17-03-94	AU-B-	4839493	29-03-94
EP-A-0215548	25-03-87	US-A- DE-A- DE-T- JP-A- US-A- US-A- US-A-	4968626 3688900 3688900 62111690 4959318 5073609 5302529	06-11-90 23-09-93 09-12-93 22-05-87 25-09-90 17-12-91 12-04-94
WO-A-9217566	15-10-92	AU-B- AU-A- JP-T-	660671 1580692 6506356	06-07-95 02-11-92 21-07-94
WO-A-9012025	18-10-90	US-A-	5043270	27-08-91
EP-A-0260148	16-03-88	AU-B- AU-B- DE-A- FR-A- GB-A, B US-A- JP-A-	613316 7831787 3730599 2603899 2197321 5024939 63152986	01-08-91 19-05-88 07-07-88 18-03-88 18-05-88 18-06-91 25-06-88
EP-A-0160457	06-11-85	AU-B- AU-B- AU-B- EP-A- HK-A- JP-A- JP-A- NO-B- SG-A- US-A-	601358 4134585 5295890 0385558 8395 60243023 6040942 174934 3994 4965199	13-09-90 24-10-85 30-08-90 05-09-90 27-01-95 03-12-85 15-02-94 25-04-94 10-06-94 23-10-90